(19) World Intellectual Property Organization International Bureau





(43) International Publication Date 16 October 2003 (16.10.2003)

PCT

(10) International Publication Number WO 03/085115 A2

(51) International Patent Classification7:

C12N 15/82

(21) International Application Number: PCT/EP03/03703

(22) International Filing Date: 8 April 2003 (08.04.2003)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data: 02447062.7 10 April 2002 (10.04.2002) EP 60/396,124 15 July 2002 (15.07.2002) US

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- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NI, NO, NZ, OM, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, Cl, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

 without international search report and to be republished upon receipt of that report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: IDENTIFICATION AND VALIDATION OF NOVEL TARGETS FOR AGROCHEMICALS

(57) Abstract: The invention relates to a method for identifying and validating plant targets for agrochemicals, comprising the steps of determining gene or protein expression profiles in function of the progression of an essential biological process in a plant, and the subsequent downregulation of expression of said gene or protein in a plant cell. More particularly, the effects of downregulation of the candidate target gene were directly monitored on plants locally infected with a vector mediating viral induced gene suppression in that infected plant area. The invention also relates to isolated plant genes encoding proteins involved in plant growth and development. The invention also relates to plants tolerant to agrochemicals such as herbicides or pesticides.

WO 03/085115
IDENTIFICATION AND VALUE AND VALU

IDENTIFICATION AND VALIDATION OF NOVEL TARGETS FOR AGROCHEMICALS

The invention relates to isolated plant genes encoding proteins essential for plant growth and development and to methods for identifying and validating these genes/proteins as target genes/proteins for agrochemicals, such as herbicides. A target for an agrochemical is a gene or a protein where the agrochemical interferes with when applied to the target organism.

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For the identification and validation of useful agrochemicals, the agrochemical industry traditionally relied on *in vivo* screening methods wherein chemical compounds were brought into direct contact with the living target organisms (e.g. plants for herbicide screening, insects for insecticide screening, etc.). However due to (i) the dramatic increase in the number of compounds that need to be screened to find a successful new agrochemical product, and (ii) the need to rely on very small quantities of compound such as are available in a combinatorial chemistry based compound libraries, and (iii) the need to identify compounds with a novel mode of action, the industry has developed a considerable interest in using more efficient and faster *in vitro* screening methods.

To render such *in vitro* screening methods more successful, it is essential to carefully select the tested target gene/proteins and/or the tested agrochemicals. It has been described that a more practical *in vitro* approach for finding new agrochemicals would involve identification of target genes/proteins against which the agrochemical compounds could possibly work. For this process identification of suitable target genes/proteins, the conventional methods make use of gene knock-outs of the target organism. Gene knock-out libraries are generally madê as a random collection of thousands of gene knock-outs. In these methods it is investigated if the gene/protein is essential for the growth and/or viability of the organism, since the knockout of an essential gene (when present in a homozygous state) leads to a lethal or otherwise detrimental effect on the organism. The indication that said gene/protein is essential to the organisms makes it a suitable target for an agrochemical. These conventional methods are still cumbersome and time consuming because of the use of gene-knockouts. Other techniques that are useful to estimate the essential character of a gene or its corresponding protein are based on the downregulation of said gene or protein for example via anti-sense expression technology (WO0107601).

To render an *in vitro* screening for agrochemicals more successful, it is essential to carefully select the tested target gene/proteins. Therefore a more practical *in vitro* approach for finding new agrochemicals could be a multistep process involving the steps of (1) identification of target genes/proteins against which the agrochemical compounds could possibly work, (2)

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Communicated by K. Isono

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WO 03/085115

validation of the candidate target gene as being an essential gene/protein for the organism and
(3) use of these target genes/proteins in an *in vitro* screening procedure in which the chemical compounds are tested.

It is the aim of the present invention to develop a process for the more efficient identification of candidate target genes/proteins for agrochemicals, combined with the more efficient validation of the target genes/proteins. It is a further aim of the invention to provide this process in order to design more efficiently the screening procedure with the agrochemical compound.

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The method of the present invention is based on the direct use of genetic information for example generated by expression profiling of the candidate target genes/proteins, for the identification and the validation of the targets.

Therefore according to a first embodiment of the present invention, there is now provided a method for identifying and validating plant genes/proteins as targets for agrochemicals, said method comprising the steps of:

- a. determining gene or protein expression profiles during a biological process of a plant or plant cell, said biological process being necessary for the viability or the growth of the plant or plant cell;
- b. selecting genes or proteins having altered expression during said biological process,
- c. cloning said selected gene or the nucleic acid encoding said protein in its full-length or partial form,
- d. incorporating said nucleic acid in a vector designed for downregulation of expression of said nucleic acid or the sequence homologous to said nucleic acid in a plant or plant cell.

The aim of methods of the present invention is the identification of target gene(s)/protein(s) out of a broad range of candidate plant genes/proteins. The identification step is achieved by the techniques of expression profiling described in the following embodiments. Since the method of the present invention can be used for identification of genes/proteins or proteins, the term "target" as used herein can mean a gene as well as a gene product, namely a protein, polypeptide or peptide. With the expression "target for an agrochemical" is meant a protein as well as a gene or nucleic acid encoding such protein, and when such target is inhibited, stimulated or otherwise disrupted in its normal activity by an agrochemical compound, this would lead to a desired effect in a target organism. The invention aims at efficiently identifying targets for agrochemicals. Said agrochemicals can be herbicides or pesticides as well as growth-stimulators or growth-regulators.

WO 03/085115

Targ et identification means selecting candidate targets from a larger number of genes/proteins or proteins on the basis of certain properties that give such a molecule a higher probability of being a suitable target than other molecules which do not exhibit said properties.

A herbicide target is a protein or gene that when inhibited, stimulated or otherwise disrupted in its normal activity by a compound would kill the (weedy) target plant or have a strong negative effect on its growth, said compound would therefore be a candidate herbicide. An insecticide target is a protein or gene that when inhibited, stimulated or otherwise disrupted in its normal activity by a compound would kill the insect pest or have a strong negative effect on its growth, said compound would therefore be a candidate insecticide. A plant growth regulator (PGR) target is a protein or gene that when inhibited, stimulated or otherwise disrupted in its normal activity by a compound would promote or alter in a desirable way the growth of plant, said compound would therefore be a candidate PGR.

Nowadays a lot of genomic information, e.g. gene sequences, expression profiles, homologies and putative functionality, is available from genomic sequencing and expression studies in several target organisms. It is therefore of interest to develop a new method to identify and validate genes/proteins as candidate targets for agrochemicals, such methods being based on a direct use of such genomic information. This use of genomic information, e.g. the expression level of a gene, allows the selection of a limited set of appropriate candidate genes/proteins. Only this limited set of genes is then tested in the validation step, contributing to a higher efficiency and success rate of the screening procedure for agrochemicals. Furthermore, the genetic information, e.g. the functional data of the putative target gene/protein, is used as a basis to design more efficiently the *in vitro* screening procedure with the agrochemical compound(s) under investigation.

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The present invention discloses methods that allow for the identification and validation of target genes/proteins for agrochemicals out of the broad range of possible genes/proteins and proteins. It therefore allows genes or proteins to be selected for the development of suitable *in vitro* screening methods for the screening of novel and efficient agrochemicals.

According to a first step of the methods of the present invention target genes or gene products are identified by using transcript profiling of the genomic content of a cell. By using this technique one immediately obtains genomic data (sequences and expression level) as well as a functional indication of the candidate target gene or gene product. Thus this method is useful for a first identification and selection of possible agrochemical target genes/proteins, since it provides as a bonus genomic and functional data on the candidate target. A good candidate target gene is a gene of which the expression varies significantly over the course of an essential biological process of the cell, since that is an indication that the gene/protein is

involved in that biological process The present application describes for the first time that the determination of an expression profile of a gene during the progression of an essential biological process is used to identify possible agrochemical targets.

The expression profiling in the target identification steps of the method of the present invention is carried out in function of the progression of a process that is essential for plant growth and/or plant development and/or plant viability. In one preferred embodiment of the present invention, the essential process that is monitored in the target identification step is the process of cell division. Accordingly, in a particular embodiment of the invention, the method to identify target genes/proteins for agrochemicals is based on the transcript profiling of genes/proteins that are specifically involved in cell division. Therefore the invention provides a method as mentioned above, wherein said biological process cell division.

Other biological processes that may be monitored for the identification and validation of agrochemical targets are for instance processes that are essential for seed germination, leaf formation, etc.

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The term expression profiling means determining the time and/or place when or where a gene or a protein is active. Particularly for a gene, this is achieved by monitoring the level of transcripts and therefore in the case of gene expression profiling the term transcript profiling or mRNA profiling is used.

Generally, the expression profiling in the methods of the present invention is carried out in function of the progression of a process that is essential for plant growth and/or development and/or plant viability. To achieve this, the process of interest is synchronized in a sufficient number of cells (for example in a cell culture) or organisms to allow collecting samples for expression profiling representing various stages of said process. Target identification then consists in selecting those genes or proteins that show significant changes in expression levels in function of the progression of the process of interest. It are those genes or proteins that are likely to be strongly involved or to be essential in said process.

The term "essential" means that if the gene or the gene product cannot function as normal in the cell or organism, this will have significant implication in the cell growth or cell development or other vital functions of the cell or organism.

According to the invention, the expression profiling can be studied at the level of m-RNA, using transcript profiling techniques, or alternatively at the level of protein, using proteomics-based approaches.

WO 03/085115

In one preferred embodiment of the invention, m-RNA profiling is used for identification of target genes/proteins and expression levels may be quantified via techniques that are well known to the man skilled in the art. For instance, mRNA-profiling can be performed using micro-array or macro-array technologies, this method however requires that the gene sequences are known (full length sequences or at least partial sequences) and are physically available for coating on the micro or macro array surface. Standard chips are being commercialised for Arabidopsis, and sufficient sequence information is now available for different plant species (including rice) to allow sufficient sequence data for this approach.

Another approach for mRNA profiling is the use of AFLP-based transcript profiling as described in example 1. In this approach short sequence tags are monitored. In a next step these short sequence tags may be matched with full-length genes/proteins if required. Gene or protein selection thus be based on either full-length or partial sequences and it is well within the realm of the person skilled in the art to find a full length sequence based on the knowledge of a partial sequence.

Therefore, one aspect of the invention is the direct use of genetic information to select 15 candidate targets for agrochemicals. As mentioned above this genetic information can be generated by a number of techniques. Accordingly, the present invention encompasses a method as mentioned above, wherein the expression profiles are determined by means of micro-array, macro array or c-DNA-AFLP.

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According to another embodiment of the invention, proteomic based approaches may be used to identify candidate target proteins for agrochemicals.

It is now demonstrated that for the purposes of identifying a target gene for agrochemicals a synchronized culture of dividing plant cells is used to isolate samples and to monitor the expression of the transcripts of those cells during the progression of the cell division.

Therefore according to a particular embodiment, the invention also encompasses a method for the identification and validation of plant agrochemical targets, wherein said gene or protein expression profiling is based on nucleic acid or protein samples collected from a synchronized culture of dividing plant cells.

In one embodiment of the invention, the samples used for expression profiling are obtained from a synchronized culture of rice cells, tobacco cells, Arabidopsis cells or cells from any other plant species. The cell culture should be synchronized in order to obtain samples containing a sufficient amount of cells that are at the same stage of the biological process, so that the various samples taken for expression profiling are representative for the various

stages of the essential biological process. In a particular embodiment of the present invention the samples are obtained from cells that are synchronized for cell division. In a preferred embodiment of the invention expression profiling is done on synchronized dividing cells. Certain cell lines are particularly suitable for synchronization of cell division, for instance synchronization of tobacco Bright Yellow-2 cell lines as described in example 1. Therefore most preferably, the synchronized cells are tobacco BY2 cells. By using synchronized tobacco BY2 cells and performing a cDNA-AFLP-based genome-wide expression analysis, the inventors built a large collection of plant cell cycle-modulated genes/proteins. Approximately 1340 periodically expressed genes/proteins were identified, including known cell cycle control genes as well as numerous novel genes. A number of plant-specific genes were found for the first time to be cell cycle modulated. Other transcript tags were derived from unknown plant genes showing homology to cell cycle-regulatory genes of other organisms. Many of the genes encode novel or uncharacterised proteins, indicating that several processes underlying cell division are still largely unknown. These sequences are presented herein as SEQ ID NO 1 to SEQ ID NO 785.

While, according to the invention, the basic criterion for identifying an agrochemical target gene or gene product consists in the differential expression levels of the gene or the protein observed during the progression of an essential biological progress, secondary selection criteria can be used and combined with this primary criterion.

One such secondary criterion may be to make a selection of genes or proteins that are found not to exhibit a high degree of homology with genes or proteins from other organisms (such as mammals) as this criterion is likely to reduce the probability that the agrochemical compounds active on the "plant-specific" target genes or gene products would also exhibit toxic effects against other organisms, for example mammals.

Another secondary selection criterion could exist in focussing on a particular phase of the essential biological process as mentioned above. For instance, when cell division modulated genes/proteins are under investigation as potential agrochemical target genes/proteins, one could preferably use those cell division modulated genes/proteins which exhibit high expression during the G1 phase, S phase, G2 phase or M phase or at the transition stages of these phases. In one embodiment of the present invention, the focus may be on the G2/M transition phase, since this phase in the plant cell cycle is considered to have more "plant specific" elements than other phases of the cell cycle and is therefore more likely to yield plant specific candidate target genes and proteins. Whereas the core cell cycle genes/proteins and the basic regulatory mechanisms controlling cell cycle progression are conserved among higher eukaryotes, basic developmental differences between plants and other organisms imply

WO 03/085115

that plant-specific regulatory pathways exist that control cell division. Especially for events occurring at mitosis, plants are expected to have developed unique mechanisms regulating karyo- and cytokinesis. A typical plant cell is surrounded by a rigid wall and can as such not divide by constriction. Instead, a new cell wall between daughter nuclei is formed by a unique cytoskeletal structure called the phragmoplast, whose position is dictated by another cytoskeletal array called the preprophase band. Another major difference between plant and animal mitosis is found in the structure of the mitotic spindles: in animals, they are tightly centred at the centrosome, whereas in plants they have a diffuse appearance.

Therefore a suitable second criterion to combine with the first criterion may be to select genes/proteins that are involved in the mitosis step of the cell cycle and/or that are involved in the building of the cell wall during mitosis.

Likewise a secondary selection criterion to be combined with the first criterion may be the selection of genes or proteins from a dicotyledonous plant that do not exhibit a high degree of homology with genes or proteins from a monocotyledonous plant (or vice versa). This secondary criterion is especially relevant when identifying agrochemical target genes or proteins with the intention to selectively identify targets that would allow for subsequence screening of selective herbicides or plant growth regulators. For instance, this strategy is advantageous to find targets and agrochemicals for selective weed control, such as herbicides that kill dicotyledonous weeds in monocotyledonous crops or vice versa.

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Therefore according to further embodiments, the present invention encompasses methods as mentioned above, wherein the target gene or protein meets any one or more of the above mentioned secondary selection criteria, such as being plant specific, being mitosis specific or being dicot specific etc.

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The possibility for combination of criteria used for selecting target genes or proteins renders the method of the present invention more powerful than classical methods. According to a preferred embodiment the technique of the present invention allows identifying genes/proteins, to be used as agrochemical target genes/proteins, these genes being genes/proteins that are involved in cell division and control of cell cycle progression, and these genes being novel and these genes being plant specific. Therefore the method of the present invention is characterized in that it allows identifying new and unexpected agrochemical targets.

In the target gene identification step according to the present invention, genes or proteins are selected for which there is a high probability of being essential. It should be clear that the above-mentioned examples are given by way of illustration and are not meant to be limiting in any way.

Further, according to a second step in the method of the invention, the candidate agrochemical target gene or gene product is subsequently validated as being essential for the growth and/or development and/or viability of the organism. This is achieved by cloning the identified candidate target gene in a vector construct designed to downregulate said target gene in a plant or plant cell, followed by inoculating the plant with this construct and monitoring whether downregulation of the gene results in negative effects on plant growth and/or development and/or viability. A valid target gene is a target gene that causes significant effects on growth of plants or plant cells when downregulated. The present application describes for the first time the use of a particularly fast and efficient downregulation method to validate possible agrochemical targets.

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Accordingly, the present invention encompasses a method as mentioned above for the identification and validation of plant targets for agrochemicals, wherein said downregulation involves a viral-induced gene silencing mechanism.

Thus, starting from a number of candidate target genes/proteins identified in the first step of the method of the invention, the target validation step aims at confirming and demonstrating the essential nature of the gene by demonstrating that severe down-regulation of the expression level of the gene has a significant effect on the organism.

In particular, when one is interested in developing a screening assay for herbicides, downregulation of the candidate target gene in a plant may result in a lethal effect, a severe inhibition of plant growth or any other (obviously) negative phenotypic effects. Alternatively, when one is interested in developing a screening assay for plant growth regulators, the effect of downregulating the target gene may be modulation or even stimulation of growth in general or modulation or even stimulation of a particular process associated with plant growth and/or development and/or architecture and/or physiology and/or biochemistry or any other phenotypic effect.

The man skilled in the art will be aware of various methods to achieve downregulation of a given gene or protein, such methods include essentially co-suppression based approaches or anti-sense based approaches as well as any other method resulting in gene silencing. Other examples of downregulation in a cell are well documented in the art and include, for example, RNAi techniques, the use of ribozymes etc. Gene silencing may also be achieved by insertion mutagenesis (for example, T-DNA insertion or transposon insertion) or by gene silencing strategies as described by, among others, Angell and Baulcombe, 1998 (WO 98/36083), Lowe et al., 1989 (WO 98/53083), Lederer et al., 1999 (WO 99/15682) or Wang et al., 1999 (WO

99/53050). Expression of an endogenous gene may also be reduced if the endogenous gene contains a mutation.

The effect of gene downregulation can be observed in stably transformed plants which can be obtained by means of various well known techniques, these techniques generally involving a plant transformation step and a plant regeneration step.

Genes/proteins which exhibit a severe negative effect when downregulated may however significantly reduce transformation and/or regeneration efficiency. Therefore, a relevant parameter indicative for the essential nature of the gene, may be a severe reduction in transformation efficiency when said particular gene is used in a down-regulation construct. In order to avoid the (negative) effect on transformation efficiency in the transformation and regeneration process, an inducible promoter system can be used. Induction of promoter activity can then be applied at a later stage (after transformation) in order to observe the effect of gene downregulation once the transformed plant or plantlet started to develop.

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Further, another method for testing the effect of downregulation of a target gene, which can be used in the methods of the present invention, is based on a rapid transient transformation process and does not rely on the somewhat lengthy process of stable transformation. The use of this method for target validation in plants is part of this invention, regardless of whether target identification has been performed according to this invention.

Accordingly, in a preferred embodiment, the downregulation method is based on co-suppression and on rapid transient transfection of plant cells. The preferred method to validate genes/proteins as targets for agrochemicals is based on the cloning of the identified candidate target gene in a vector construct containing a viral replicase that is involved in the very efficient downregulation of the candidate target gene in the infected plant or plant cell via the mechanism of co-suppression. One advantage of this method for downregulation, is the fact that the infection of the host cells or the plant can be performed locally for example by inoculating the vector directly on the leaves. This allows a very fast evaluation of the effect of downregulating the candidate target since no complete transgenic plants have to be generated. Also this technique allows an easy way of monitoring the effect of the downregulated candidate target by simply looking at the changes of the infected place, for example monitoring the lethal effects on the infected leaf).

Therefore in a preferred embodiment, the downregulation method is based on co-suppression. In a more preferred embodiment of the invention this co-suppression technique is fast and easy to evaluate the effect of downregulation, so that it is suitable for dealing with high

numbers of genes/proteins. This can be achieved by using viral induces gene silencing mechanisms (VIGS) and by infecting the plant directly and locally, for example on the leaves. Therefore, according to another embodiment, the present invention relates to the use of a viral-induced gene silencing system for validating plant targets for agrochemicals.

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This method for severe downregulation via transient expression of the gene in the presence of certain viral elements is referred to as "virus-induced gene silencing mechanism" (VIGS) and is previously described in Ratcliff *et al.*, Plant J., 25 237 – 245, 2001. Briefly, virus vectors carrying host-derived sequence inserts induce silencing of the corresponding genes/proteins in infected plants. This virus-induced gene silencing is a manifestation of an RNA-mediated defence mechanism that is related to post-transcriptional gene silencing in transgenic plants. Ratcliff *et al.*, developed an infectious cDNA clone of Tobacco rattle virus (TRV) that has been modified to facilitate insertion of non-viral sequences and subsequent infection in plants. This vector mediates VIGS of endogenous genes/proteins in the absence of virus-induced symptoms. Unlike the other RNA virus vectors that have been used previously for VIGS, the TRV construct is able to target most RNA's in the growing points of the plant. A more detailed description of this downregulation mechanism is given in example 2.

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gene.

According to a further preferred embodiment, there is provided a method for validation of a candidate agriochemical target gene, wherein the gene is downregulated in a plant via the use

of infectious DNA of virus is Tobacco Rattle Virus and wherein said plant is tobacco.

According to particular embodiments of the present invention, the VIGS system is applied in

Arabidopsis or in tobacco for the purposes of validation of a candidate agrochemical target

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The present invention relates to a combination of the above-mentioned identification and validation steps, which are especially selected so that they lead to an efficient selection of candidate target genes for agrochemicals. The outcome of the transcript profiling provides the necessary information and forms the basis for the second step, namely the validation of the target gene via incorporation of the gene sequence in the downregulation construct. The combination of these two techniques is especially useful for selecting suitable target genes/proteins for agrochemicals in a high throughput fashion. This technique thus overcomes the technical limitations of previously described techniques such as the knock-out libraries and the antisense strategies without genetic information of the genes. This new combination offers a time-saving strategy for identification of a candidate target gene and the more direct information output in the form of a real sequence, the immediate cloning of the gene in the

WO 03/085115

downregulation construct and immediate application of the downregulating construct on the target organism.

The combination of these steps offers the unique opportunity to provide many high quality target genes/proteins for agrochemicals in a commercially and economically advantageous way. Furthermore, inherent to the techniques of the present invention is that the qualified target genes/proteins are accompanied with the necessary information to design a suitable in vitro screening assay with the agrochemical. This information consists of the expression characteristics of the genes/proteins and their function and importance in the essential biological process that was monitored during the transcript profiling.

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In this way, the methods of the present invention overcome the practical and commercial 10 limitations of the existing techniques.

Once this level of target validation is reached, the validated target can be selected for the development of an appropriate high-throughput in vitro screening method, wherein the agrochemical is tested. Therefore, the present invention also encompasses a method for screening candidate agrochemical compounds, comprising the use of any of the identification procedures and/or validation procedures as mentioned above. More particularly, the present invention encompasses a method for screening agrochemical compounds, comprising the use of any one or more of the sequences represented in SEQ ID NO 1 to 785.

Various methods can be used to develop suitable in vitro assays for screening the chemical 20 compounds, depending on what is known about the biological activity of the target gene. For example, when the target is an enzyme, measurement of the enzymatic activity of the target could form the basis of the in vitro screening assay with the chemical compound.

Therefore, the methods of the present invention, the genes/proteins and the information 25 generated by the combined identification and validation methods of the present invention, allow one to design and/or fine tune a screening for testing and/or developing agrochemicals (for example herbicides). For example if the expression pattern and the role of the target gene in the essential biological process is known, it is much easier to set up an in vitro screening assay to monitor the effect of a candidate herbicide on the target cells. Therefore it is expected 30 that much more refined and/or efficient herbicides will be characterized using the methods of

Also because of the knowledge of its function, one can further design the screened agrochemical compound to improve its activity for instance to improve its binding capacity to

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Therefore, the present invention encompasses a method for screening candidate agrochemical compounds comprising the use of any of the methods as mentioned above.

The invention may also be applied for the development of agrochemical (for example herbicide or pesticide) tolerant plants, plant tissues, plant seeds and plant cells.

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Herbicides that exhibit greater potency can also have greater crop phytotoxicity. A solution to this problem is to develop crops that are resistant or tolerant to herbicides. Crop hybrids or varieties that are tolerant to the herbicides allow, for instance, for the use of herbicides that kill weeds without attendant risk of damaging the crop. Further it should be clear that when a plant is overexpressing the target of a particular herbicide, the tolerance of said plant against said herbicide will also be enhanced.

Therefore the present invention also relates to the use of the agrochemical (e.g. herbicide) target genes/proteins as identified by the method of the present invention for generating transgenic plants that are tolerant or resistant to an agrochemical (e.g. herbicide). Example of genes and gene sequences identified by the combined identification and validation methods of the present invention and which can be used as agrochemical target or that can be used to obtain herbicide tolerant plants comprise the sequences as represented in any of SEQ ID NOs 1 to 785.

These sequences are derived from tobacco, but the one skilled in the art can easily find via homology search in databases or homology search in a cDNA library the homologues genes of other plant species, for instance monocot sequences (e.g. the corresponding rice or corn sequence), and use them for the same purposes as described herein. These homology searches can be done for example with a BLAST program (Altschul et al., Nucl. Acids Res., 25 3389 – 3402, 1997) on a sequence database such as the GenBank database. Homology studies as referred to above can be performed using sequences present in public and/or proprietary databases and using several bioinformatics algorithms, well known to the man skilled in the art. Methods for the alignment of sequences are well known in the art, such methods include GAP, BESTFIT, BLAST, FASTA and TFASTA. GAP uses the algorithm of Needleman and Wunsch (J. Mol. Biol. 48: 443-453, 1970) to find the alignment of two complete sequences that maximizes the number of matches and minimizes the number of gaps. The BLAST algorithm calculates percent sequence identity and performs a statistical analysis of the similarity between the two sequences. The software for performing BLAST analysis is publicly available through the National Centre for Biotechnology Information.

Further, some of the tobacco sequences identified by the method of the present invention might be partial but again, the full-length sequence can easily be found based on the partial

sequence. For example "transcript building" can be done based on homology search on EST databases, cDNA's or gene predictions. These databases and programs are publicly available e.g. http://www.tigr.org/.

Therefore the present invention relates to the use of the nucleic acids as identified and disclosed herein and represented in SEQ ID NO 1 to 785, and also to the use of the full length genes regenerated from the partial sequences as well as to the use of the homologues sequences isolated from the same or from other plants.

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In another embodiment, the present invention relates to a nucleic acid identified according to the method of the invention. Thus the invention encompasses an isolated nucleic acid identifiable by any of the methods as mentioned above.

In another embodiment, the invention relates to a nucleic acid identified according to the method of the invention, comprising the nucleic acid sequence chosen from the group of SEQ ID NO 1 to 785 or a full length sequence thereof, or a functional homologue thereof, or a functional fragment thereof, or an immunologically active fragment thereof. Thus the invention encompasses an isolated nucleic acid, comprising at least part of a nucleic acid sequence chosen from the group of SEQ ID NO 1 to 785 a homologue, functional fragment or derivative thereof.

With "a functional fragment" is meant any part of the sequence that is responsible for the biological function or for an aspect of the biological function of the nucleic acid sequence.

Further, the invention encompasses a method for the production of an agrochemical resistant plant, comprising the use of any one or more of SEQ ID NO 1 to 785 or a homologue, functional fragment or derivative thereof or one or more of the proteins encoded by SEQ ID NO 1 to 785 or a homologue, functional fragment or derivative thereof.

In one embodiment of the present invention the sequences, the full-length sequences and the homologues are used to develop herbicide tolerant plants.

Further the invention encompasses a plant tolerant to an agrochemical, in which the expression level of one or more of the nucleic acids corresponding the SEQ ID NO 1 to 785 or the homologue, functional fragment or derivative thereof, is modulated. Further the invention encompasses any part or more preferably any harvestable part of these plants.

Therefore the invention also relates to the use of these sequences, the full-length sequences and the homologues as targets for agrochemicals The invention encompasses the use of a

nucleic acid as mentioned above or the protein encoded by said isolated nucleic acid as a target for an agrochemical compound, preferably, wherein the agrochemical compound is a herbicide.

5 Further, the invention relates to the use of these sequences to develop screening assays for the identification and/or development of agrochemicals. The invention encompasses a method for screening candidate agrochemical compounds comprising the use of any one or more of SEQ ID NO 1 to 785 or a homologue, functional fragment or derivative thereof or one or more of the proteins corresponding to SEQ ID NO 1 to 785 or a homologue, functional fragment or derivative thereof.

The present-invention will be further illustrated by the following figures, wherein,

Figure 1 shows the gene expression profiles obtained by quality-based clustering of all transcript tags monitored in a transcript profiling experiment as described in example 1. Shown are the trend lines of 16 clusters containing 97% of the genes and covering the entire time course as indicated on top. S-phase-specific gene clusters are grouped in A, gene clusters with peak expression between S- and M-phase are grouped in B, whereas group C contains the M- and G1-phase-specific clusters. D: Three small clusters of genes with peak expression during two cell cycle phases.

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Figure 2 shows the phenotypes of tobacco plants inoculated with a acetolactate synthase (SEQ ID NO 18) downregulation construct and phenotypes of tobacco plants inoculated with a prohibitin (SEQ ID NO 21) downregulation construct. The phenotypes were observed 12 days after inoculation (upper panel) or 17 days after inoculation (lower panel).

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Figure 3 shows the phenotype of tobacco plants inoculated with a B-type CDK (SEQ ID NO 11) donwregulation contruct. The observations were made 37 days after inoculation.

Figure 4 shows the sequences identified by the methods of the present invention and represented by SEQ ID NO 1 to SEQ ID NO 785

EXAMPLES

Example 1

A cDNA-AFLP based expression profiling of sequence obtained from samples of a synchronized tobacco BY2 cell line system, was used to identify genes that are upregulated during the cell cycle, an essential biological process needed for the viability and growth of the tobacco cell line system.

A genome-wide expression analysis of cell cycle-modulated genes in the tobacco Bright Yellow-2 (BY2) cell line was performed. This unique cell line can be synchronized to high levels with different types of inhibitors of cell cycle progression (Nagata *et al.*, Int. Rev. Cytol., 132 1 – 30, 1992; Planchais *et al.*, FEBS Lett., 476 78 –83, 2000). Because of the lack of extensive molecular resources such as genomic sequences, cDNA clones or expressed sequence tags (ESTs) for tobacco, a microarray-based approach cannot be used for a transcriptome analysis. Therefore, the cDNA-AFLP technology was used to identify and characterize cell cycle-modulated genes in BY2. cDNA-AFLP is a sensitive and reproducible fragment-based technology that has a number of advantages over other methods for genome-wide expression analysis (Breyne and Zabeau, Curr. Opin. Plant Biol., 4 136 – 142, 2001): it does not require prior sequence information, it allows identification of novel genes, and it provides quantitative expression profiles. After a detailed analysis, it was found that around 10% of the transcripts analyzed is periodically expressed. This comprehensive collection of plant cell cycle-modulated genes provides a basis for selecting and validating novel and unexpected agrochemical target genes

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Synchronization of BY2 cells and sampling of material. Tobacco BY2 (Nicotiana tabacum L. cv. Bright Yellow-2) cultured cell suspension were synchronized by blocking cells in early S-phase with aphidicolin as follows. Cultured cell suspension of Nicotiana tabacum L. cv. Bright Yellow 2 were maintained as described (Nagata et al., Int. Rev. Cytol., 132 1 - 30, 1992). For synchronization, a 7-day-old stationary culture was diluted 10-fold in fresh medium supplemented with aphidicolin (Sigma-Aldrich, St. Louis, MO; 5 mg/l), a DNA-polymerase α inhibiting drug. After 24 h, cells were released from the block by several washings with fresh medium and resumed their cell cycle progression. After the drug had been washed, samples were taken every hour, starting from the release from the aphidicolin block (time 0) until 11 h later. The mitotic index was determined by counting the number of cells undergoing mitosis fluorescence microscopy after the DNA had been stained with 5 mg/l 4',6-diamidino-2-phenylindole (Sigma-Aldrich). DNA content was measured by flow cytometry. This was done as follows A subsample was used to check cell cycle progression and synchrony levels. After the DNA had been stained with 5 mg/l 4',6-diamidino-2-phenylindole (Sigma-Aldrich), the mitotic index was determined under fluorescence microscopy by counting the number of cells undergoing mitosis. A mitotic peak of approximately 40% was obtained 8 h after washing. For flow cytometry, cells were first incubated in a buffered enzyme solution (2% cellulase and 0.1% pectolyase in 0.66 M sorbitol) for 20 min at 37°C. After the suspension had been washed and resuspended in Galbraith buffer (Galbraith et al., Science, 220 1049 - 1051, 1983), it was filtered through a 30- μm nylon mesh to purify the DAPI-stained nuclei. The fluorescence intensity was measured using a BRYTE HS flow cytometer (Bio-Rad, Hercules,

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CA). Exit from S-phase was observed 4 h after aphidicolin release and the level of synchrony was shown to be sufficiently high throughout the time course.

RNA extraction and cDNA synthesis. Total RNA was prepared by using LiCI precipitation (Sambrook et al., 1989) and poly(A⁺) RNA was extracted from 500 µg of total RNA using Oligotex columns (Qiagen, Hilden, Germany) according to the manufacturer's instructions. Starting from 1 µg of poly(A⁺) RNA, first-strand cDNA was synthesized by reverse transcription with a biotinylated oligo-dT₂₅ primer (Genset, Paris, France) and Superscript II (Life Technologies, Gaithersburg, MD). Second-strand synthesis was done by strand displacement with *Escherichia coli* ligase (Life Technologies), DNA polymerase I (USB, Cleveland, OH) and RNAse-H (USB).

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cDNA-AFLP analysis. Five hundred ng of double-stranded cDNA was used for AFLP analysis as described (Vos et al., Nucl. Acids Res., 23 4407 - 4414, 1995; Bachem et al., Plant J., 9 745 - 753, 1996) with modifications. The restriction enzymes used were BstYI and Msel (Biolabs) and the digestion was done in two separate steps. After the first restriction digest with one of the enzymes, the 3' end fragments were collected on Dyna beads (Dynal, Oslo, Norway) by means of their biotinylated tail, while the other fragments were washed away. After digestion with the second enzyme, the released restriction fragments were collected and used as templates in the subsequent AFLP steps. The adapters used were: for BstYl, 5'-CTCGTAGACTGCGTAGT-3' and 5'-GATCACTACGCAGTCTAC-3', and for Msel, 5'-GACGATGAGTCCTGAG-3' and Msel were 5'-TACTCAGGACTCAT-3'; primers for BstYl and the 5'-GACTGCGTAGTGATC(T/C)N₁₋₂-3' and 5'- GATGAGTCCTGAGTAAN₁₋₂-3', respectively. For preamplifications, a Msel primer without selective nucleotides was combined with a BsfYl primer containing either a T or a C as 3' most nucleotide. PCR conditions were as described Vos et al., Nucl. Acids Res., 23 4407 - 4414, 1995). The obtained amplification mixtures were diluted 600-fold and 5 ul was used for selective amplifications using a P³³-labeled BsfYI primer and the Amplitaq-Gold polymerase (Roche Diagnostics, Brussels, Belgium). Amplification products were separated on 5% polyacrylamide gels using the Sequigel system (Biorad). Dried gels were exposed to Kodak Biomax films as well as scanned in a phospholmager (Amersham Pharmacia Biotech, Little Chalfont, UK).

Quantitative measurements of the expression profiles and data analysis. Gel images were analyzed quantitatively with the AFLP-QuantarPro image analysis software (Keygene N.V., Wageningen, The Netherlands). This software was designed for accurate lane definition, fragment detection, and quantification of band intensities. All visible AFLP fragments were scored and individual band intensities were measured per lane. The obtained data were used to

determine the quantitative expression profile of each transcript. The raw data were corrected for differences in total lane intensities, after which each individual gene expression profile was variance-normalized. This was done as follows.

The obtained raw data were first corrected for differences in total lane intensities which may occur due to loading errors or differences in the efficiency of PCR amplification with a given primer combination for one or more time points. The correction factors were calculated based on constant bands throughout the time course. For each primer combination, a minimum of 10 invariable bands was selected and the intensity values were summed per lane. Each of the summed values was divided by the maximal summed value to give the correction factors. Finally, all raw values generated by QuantarPro were divided by these correction factors.

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Subsequently, each individual gene expression profile was variance-normalized by standard statistical approaches as used for microarray-derived data (Tavazoie *et al.*, Nature Genet., 22 281 – 285, 1999). For each transcript, the mean expression value across the time course was subtracted from each individual data point after which the obtained value was divided by the standard deviation. A coefficient of variation (CV) was calculated by dividing the standard deviation by the mean. This CV was used to establish a cut-off value and all expression profiles with a CV less than 0.25 were considered as constitutive throughout the time course.

The Cluster and TreeView software (Eisen *et al.*, PNAS, 95 14863 – 14868, 1998) was used for hierarchical, average linkage clustering. Quality-based clustering was done with a newly developed software program (De Smet et al., Bioinformatics 2002 May; 18(5): 735-46). This program is related to K-means clustering, except that the number of clusters does not need to be defined in advance and that the expression profiles that do not fit in any cluster are rejected. The minimal number of tags in a cluster and the required probability of genes belonging to a cluster were set to 10 and 0.95, respectively. With these parameters, 86% of all the tags were grouped in 21 distinct clusters.

Characterization of AFLP fragments. Bands corresponding to differentially expressed transcripts were isolated from the gel and eluted DNA was reamplified under the same conditions as for selective amplification. Sequence information was obtained either by direct sequencing of the reamplified polymerase chain reaction product with the selective *Bst*YI primer or after cloning the fragments in pGEM-T easy (Promega, Madison, WI) or sequencing of individual clones. The obtained sequences were compared against nucleotide and protein sequences present in the publicly available databases by BLAST sequence alignments (Altschul *et al.*, Nucl. Acids Res., 25 3389 – 3402, 1997). When available, tag sequences were replaced with longer EST or isolated cDNA sequences to increase the chance of finding significant homology. Based on the homology, transcript tags were classified in functional groups as shown in Table 1.

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Experimental Results

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Identification and characterization of cell cycle-modulated genes

Tobacco BY2 cells were synchronized by blocking cells in early S-phase with aphidicolin, an inhibitor of DNA polymerase α . After the inhibitor had been released, 12 time points with an 1-h interval were sampled, covering the cell cycle from S-phase until M-to-G1 transition. Flow cytometry and determination of the mitotic index showed that the majority of cells exit S-phase 4 h after release from blocking and that the peak of mitosis is reached at 8 h. From each time point, extracted mRNA was subjected to cDNA-AFLP-based transcript profiling.

Quantitative temporal accumulation patterns of approximately 10,000 transcript tags were determined and analyzed. In total, around 1,340 transcript tags were modulated significantly during the cell cycle: Hierarchical clustering of the expression profiles resulted in four large groups with the peak of expression in S-, early G2-, late G2-, or M-phase. Within each of these groups, several smaller clusters of genes with similar expression patterns could be distinguished. By quality-based clustering 21 different clusters were identified (see: http://www.plantgenetics/genomics/CCMgenes). In agreement with the hierarchical clustering, the four largest clusters (clusters 1 to 4 in Fig. 1) correspond to the S-, early G2-, late G2-, and M-phases and together contain 65% of all the tags. An additional cluster (cluster 5 in Fig. 1C), not clearly separated in the hierarchical clustering, includes the genes with peak expression in G1-phase and contains another 5% of the tags. The remaining clusters are much smaller and most often (e.g., clusters 6, 9, 10, and 18) include genes with a narrow temporal expression pattern. In addition to these clusters, three small groups of genes displaying elevated expression during two cell cycle phases were distinguished also by quality-based clustering (Fig. 1 D).

After the transcript tags had been sequenced, homology searches revealed that 36.5% of the tags were significantly homologous to genes of known functions, 13.1% of the tags matched a cDNA or genomic sequence without allocated function, whereas for 50.4% of the tags no homology with a known sequence was found. Genes of known function belong to diverse functional classes (Table 1) revealing that several biological processes are at least partially under temporal transcriptional control during the cell cycle in plants. In general, the observed transcript accumulation profiles and cell cycle specificity correlate well with the functional properties of the corresponding genes. It is interesting that the number of transcription factors with G2-phase specificity is high, which may be related with the induction of genes involved in M-phase-specific processes. The overrepresentation of RNA-processing genes in the M-phase might indicate that post-transcriptional regulation is involved in gene activity during mitosis. Because *de novo* transcription is severely reduced during mitosis (Gottesfeld *et al.*, Trends Bioch. Sci., 22 197 – 202, 1997). RNA-processing could provide an alternative regulatory

mechanism. Intriguingly, transcript tags with homology to a gene of unknown function are overrepresented in the M-phase as well (Table 1). The principal differences in cell cycle events between plants and other organisms occur during mitosis; therefore, the inventors believe that several of these transcripts correspond to still uncharacterised plant-specific genes triggering these events. Remarkably, several of the tags homologous to a publicly available sequence have no *Arabidopsis* homologue, indicating that, in addition to conserved genes, different plant species possess also unique sets of cell cycle-modulated genes. Although many of these tags may be too short to significantly match with an *Arabidopsis* sequence, analysis of longer cDNA clones corresponding to a subset of tags has revealed that approximately 25% of the sequences remain novel.

In Tables 1 to 4 a selection of 785 sequence tags are shown. This selection was based on the criterion if the tags were full length or that showed homology with genes known to be involved in the cell cycle (group 2 SEQ ID NOs 22 to 118), or on the criterion that they show homology with genes of unknown function (group 3 SEQ ID NOs 119 to 283) or on the criterion that the sequences showed no homology with the sequences in that existing databases (group 4 SEQ ID NOs 284-785). A first group (SEQ ID Nos 1 to 21) represent a smaller selection of tags which are used in the target validation method described in the present invention, more particularly, that were used in example 2.

20 The core cell cycle machinery

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Several tags coincide with genes belonging to the core cell cycle machinery and exhibiting distinct expression profiles. Transcript tags from five B1- or B2-type cyclins as well as from a D2-type cyclin show mitotic accumulation and exhibit a narrow temporal expression profile, confirming previous studies (Mironov et al., Plant Cell, 11 509 - 521, 1999; Sorrell et al., Plant Physiol., 119 343 - 351, 1999). Based on the transcription patterns, the six A-type cyclins fall into three groups that sequentially appear during the cell cycle, adding new data to earlier observations (Reichheld et al., PNAS, 93 13819 - 13824, 1996). Two groups have quite a broad window of transcript accumulation; one group, homologous to A3-type cyclins, is expressed during S-phase and disappears during G2-phase and the other group, corresponding to A2-type cyclins comes up at mid S-phase and goes down during M-phase, except for one transcript that is specific for S-phase. The third group, containing an A1-type cyclin, has the same expression pattern as the B- and D2-type cyclins. Several tags derived from genes encoding the plant-specific B-type cyclin-dependent kinases (CDKs) were also identified. CDKB1 and CDKB2 peak at the G2-to-M transition, slightly before the mitotic cyclins as describe (Porceddu et al., J. Biol. Chem., 276 36354 - 36360, 2001). In contrast to what has been observed in partially synchronized alfalfa cell cultures (Magyar et al., Plant Cell, 9 223 - 235, 1997), the transcript levels of the tags homologous to a C-type CDK accumulate

differentially during the cell cycle. The transcripts are present during late M-phase and early S-phase, suggesting that CDKC is active during the G1-phase.

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In addition to these well-characterized cell cycle-regulatory genes, also several tags were identified herein derived from genes encoding transcription factors and protein kinases or phosphatases with a known or putative role in cell cycle control. One tag with a sharp peak of transcript accumulation 1 h before the B- and D-type cyclins corresponds to a 3R-MYB transcription factor. Recently, a 3R-MYB has been shown to activate B-type cyclins and other genes with a so-called M-phase-specific activator domain (Ito et al., Plant Cell, 13 1891 -1905, 2001). Another tag peaking in M-phase is homologous to the CCR4 associated protein CAF. CAF forms a complex with CCR4 and DBF2, resulting in a transcriptional activator involved in the regulation of diverse processes including cell wall integrity, methionine biosynthesis and M-to-G1 transition (Liu et al., EMBO J., 16 5289 - 5298, 1997). A majority of the tags with similarity to protein kinases and phosphatases show M-phase-specific accumulation (Table 1). Although the true identity and putative cell cycle related function remains unclear for the majority, one is highly homologous to a dual-specificity phosphatase. This type of phosphatases plays a crucial role in cell cycle control in yeast and animals (Coleman and Dunphy, Curr. Opin. Cell Biol., 6 877 - 882, 1994). Another M-phase-specific tag is homologous to prohibitin. In the mammalian cell cycle, prohibitin represses E2F-mediated transcription via interaction with retinoblastoma (Rb), thereby blocking cellular proliferation (Wang et al., Oncogene, 18 3501 - 3510, 1999).

Protein degradation by the ubiquitin-proteasome pathway also plays an important role in the control of cell cycle progression at both G1-to-S transition and exit from mitosis. Although there is little evidence for cell cycle-modulated expression of the genes encoding the various components of the ubiquitin-proteasome complexes, some proteins accumulate in a cell cycle-dependent way (del Pozo and Estelle, Plant Mol. Biol., 44 123 – 128, 2000). Furthermore, several tags were isolated herein from genes encoding ubiquitin-conjugating enzyme (E3), ubiquitin-protein ligase (E2), and proteasome components with an M-phase-specific expression pattern. Another transcript tag that accumulates during late M-phase is similar to cathepsin B-like proteins, which are proteolytically active and degrade diverse nuclear proteins, including Rb (Fu et al., FEBS Lett., 421 89 – 93, 1998).

Whereas all the core cell cycle regulatory genes have been identified that control the G2-to-M transition for which the expression is known to be cell cycle modulated, genes such as Rb and E2F, controlling G1-to-S transition were not found. These genes were probably missed because the G1-to-S transition was not included in the present analysis, what is supported by the finding that the early targets of E2F, such as polymerase α and ribonucleotide reductase, are already present at high levels at the beginning of the time course.

Genes involved in DNA replication and modification

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In agreement with the studies performed in yeast and human fibroblasts, transcripts encoding proteins involved in DNA replication and modification accumulated during S-phase and exhibited broad temporal expression profiles. Different replication factors, DNA polymerase α , and the histones H3 and H4 are already present at the onset of the time course, indicating that they are induced before the time point of the aphidicolin arrest. Interestingly, most of the histones H1, H2A, and H2B appear somewhat later than H3 and H4, what might reflect that they are deposited into the nucleosomes after H3 and H4 (Luger et al., Nature, 389 251- 260, 1997; Tyler et al., Nature, 402 555 - 560, 1999). The profile of the homologue of the anti-silencing function 1 (ASF1) protein is similar to that of the histones H3 and H4, in agreement with the fact that the three proteins are part of the replication-coupling assembly factor complex that mediates chromatin assembly (Tyler et al., Nature, 402 555 ~ 560, 1999). Genes encoding high-mobility group proteins reach the highest accumulation during late G2, consistent with the subsequent steps involved in the folding and structuring of the chromatin. Tags derived from genes encoding proteins involved in DNA modification, such as S-adenosyl-L-methionine (SAM) synthase and cytosine-5-methyl- transferase are found in the histone cluster. Tags from methionine synthase genes, which provide the precursor for SAM synthase, accumulate during M-phase, in contrast to yeast, where these genes are expressed during late S-phase (Spellman et al., Mol. Cell Biol., 9 3273 - 3297, 1998).

20 Genes involved in chromatin remodelling and transcriptional activation or repression have been identified as well. One gene is a histone deacetylase with highest transcript accumulation during the G2-phase and another belongs to the SNF2 family of chromodomain proteins with an M-phase-specific expression pattern. Interestingly, one tag corresponds to a mammalian inhibitor of growth 1 (p33-ING1) protein. The human ING1 protein has DNA-binding activity and might be involved in chromatin-mediated transcriptional regulation (Cheung and Li, Exp. 25 Cell Res., 268 1 - 6, 2001). This protein accumulates during S-phase (Garkavtsev and Riabowel, Mol. Cell Biol., 17 2014 - 2019, 1997), what is in agreement with the expression profile we observed. The yeast homologues of ING1 are components of the histone acetyltransferase complex and show similarity to the Rb-binding protein 2 (Loewith et al., Mol. Cell Biol., 20 3807 - 3816, 2000). Another tag, homologous to the Arabidopsis MSI3 protein, 30 follows a similar expression profile. MSI-like proteins are involved in the regulation of histone acetylation and deacetylation and in chromatin formation (Ach et al., Plant Cell, 9 1595 – 1606, 1997).

The expression profiles of the different ribonucleotide reductase (RNR) genes are more complex. One gene is already expressed at high levels at the beginning of the time course and its expression is restricted to the S-phase as described (Chabouté *et al.*, Plant Mol. Biol., 38 797 – 806, 1998), whereas, in contrast, another one is highly expressed in S-phase and

reappears at lower levels during M-phase and a third one is M-phase-specific. This latter expression profile has also been described for a *RNR* gene from *Xenopus* where the encoded protein appears to be involved in microtubulin nucleation (Takada *et al.*, Mol. Cell Biol., 11 4173 – 4187, 2000).

Numerous other transcript tags with S-phase specificity were found in addition to the ones involved in DNA replication and modification. Most interestingly, one of these tags is homologous to a mammalian gene encoding a TRAF-interacting protein (TRIP), which is a component of the tumor necrosis factor (TNF) signalling complex, and promotes cell death when complexed with TRAF (Lee *et al.*, J. Exp. Medicine, 185 1275 – 1285, 1997). Another S-phase-specific tag shows homology to the RING finger domain of inhibitor of apoptosis proteins, which are also involved in the TNF signalling pathway.

Modulated expression of genes required for mitosis and cytokinesis

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Several paralogous genes that encode either α - or β -tubulin were highly induced and accumulated prior to the mitotic index peak or during early M-phase. The inventors found that in BY2, tubulin genes are highly cell cycle modulated. This transcriptional regulation is in agreement with previous demonstrations of de novo transcription of α - and β -tubulin genes during different cellular processes (Stotz et al., Plant Mol. Biol., 41 601 - 614, 1999). In the present analysis, no γ -tubulin genes were found, confirming published data that the amount of γ -tubulin is constant in dividing BY2 cells (Stoppin-Mellet *et al.*, Plant Biol., 2 290 – 296, 2000). Most of the kinesins identified herein, fall in the same cluster as the tubulins peaking prior to mitosis. Interestingly, two tags have a distinct transcription pattern and appear in another gene cluster. Their window of transcript accumulation is very narrow and coincides with the peak of the plant-specific these correspond to interestingly, tags mitosis. Most phragmoplast-associated type of kinesin, PAKRP1 (Lee and Liu, Curr. Biol., 10 797 - 800, 2000). A chromokinesin not yet described in plants was identified as well. This type of motor proteins use DNA as cargo and play a role in chromosome segregation and metaphase alignment (Wang et al., J. Cell Biol., 128 761 - 768, 1995).

Among the M-phase-specific kinases, two were unambiguously recognized herein as playing a role in cytokinesis. One is Aurora, a protein kinase with a key role in the control of chromosome segregation, centrosome separation, and cytokinesis in yeast and animals (Bischoff and Plowman, Trends Cell Biol., 9 454 – 459, 1999) but not described in plants yet. The other is NRK1, a mitogen-activated protein kinase kinase which is phosphorylated by NPK1, a kinase involved in regulating the outward redistribution of phragmoplast microtubules (Nishihama et al., Genes Dev., 15 352 – 363, 2001).

Hormonal regulation and cell cycle-modulated gene expression

A number of genes belonging to the class of auxin-induced genes were also differentially expressed. Cell cycle-modulated expression of auxin-induced genes has never been observed before although auxins together with cytokinins are the two major groups of plant hormones that affect cell division (Stals and Inzé, Trends Plant Sci., 6 359 – 364, 2001). The genes as identified herein fall into two groups based on their transcript accumulation profiles (data not shown). The first group displays an early S-phase-specific expression pattern and consists of the parA, parB and parC genes. Induction of the par genes is most often observed in response to stress conditions (Abel & Theologis, Plant Phys. 111, 9 – 17, 1996). The fact that the transcripts rapidly disappear after release from the cell cycle-blocking agent might indicate a stress response rather than a cell cycle dependent auxin response.

More interesting is the second group of genes with transcripts accumulating during early M-phase. This group includes the auxin response factor 1 (*ARF1*), an auxin transporter as well as different members of the early auxin response *AUX/IAA* gene family. ARF1 is a transcription factor that binds to a particular auxin response element (Ulmasov *et al.*, Science, 276 1865 – 1868, 1997). Additional studies suggest that the activity of ARF1 is controlled by its dimerization with members of the AUX1/IAA family (Walker and Estelle, Curr. Opin. Plant boil., 1 434 – 439,1998). The similarity in temporal expression profiles the inventors observed supports these findings and suggests that these proteins mediate an auxin response necessary for cell cycle progression

By using tobacco BY2 as model system together with cDNA-AFLP-based transcript profiling, it is described herein for the first time how a comprehensive inventory of plant cell cycle-modulated genes can be made. Although the obtained data confirm earlier results and observations, in addition, numerous novel findings were made. The obtained data are a very useful basis for selecting and validating agrochemical target genes.

Example 2

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In this example it is described how plant genes are evaluated for assessment of their essential character in the biological process, thus how they are validated as good candidate targets for agrochemicals.

The Tobacco Rattle Virus (TVR) is used to induce silencing of target genes. In case of an essential gene the simlencing will result in a lethal effect on the plant and therefore, the suystem allows to validate good candidates as targets for herbicides.

The TRV based system is used in this example in combination with series of candidate genes, more particularly with the candidate targets as represented herein as group 1 sequences consisting of the SEQ ID NOs 1 to 21. The identification technique of the present invention (see example 1) allowed to identify new genes that are potential new herbicide targets,

because of their putative function in various key processes crucial for cell life, their expression at a certain developmental stage crucial for cell life, their role in metabolism and/or maintenance of cell living state.

This example illustrates the validation of these candidate genes as novel targets for agrochemicals, via the technique of the virus-induced gene silencing (VIGS).

Gene silencing mechanism

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The virus-induced gene silencing (VIGS) is a manifestation of an RNA-mediated defence mechanism that is related to post-transcriptional gene silencing (PTGS) in transgenic plants (Ratcliff *et al.*, Plant J., 25 237 – 245, 2001). The method uses a vector with an infectious cDNA of tobacco rattle virus (TRV) modified (see below) to facilitate insertion of target sequences and modified for efficient infection of plants (e. g. tobacco). The vector mediates VIGS of endogenous genes in the absence of specific virus-induced symptoms.

The RNA-mediated defence is triggered by the virus vectors, and targets both the viral genome and the host gene corresponding to the insert. As a result, the symptoms in the infected plant are similar to loss-of-function mutants or reduced-expression mutants in the host gene. The presence of a negative growth phenotype suggests that the targeted gene is a potential herbicide target.

The process of constructing a virus vector and monitoring symptoms on infected plants is completed within a few weeks, such that virus-induced gene silencing (VIGS) provides a simple, rapid means of assigning function to genes that have been sequenced but are otherwise uncharacterized. The determination of new herbicide target genes is performed in a few weeks including gene cloning, transformation steps and tobacco plant analyses.

The TRV construct is shown to target host RNAs in the growing points of plants (Ratcliff *et al.*, Plant J., 25 237 – 245, 2001) such as meristems and actively dividing cells.

It has been shown that this vector overcomes many of the problem features of PVX, TMV and TGMV. For example, the TRV vector induces very mild symptoms, infects large areas of adjacent cells and silences gene expression in growing points such as meristems and actively dividing cells. Infection of tobacco plants on the leaves with TRV based constructs will affect growth and development of upper parts of the infected leaves and allow screening for growth parameters.

Construction of TRV vectors used in the validation process of the present invention

TRV is a positive-strand RNA virus with a bipartite genome. Proteins encoded by RNA 1 are sufficient for replication and movement within the host plant, while proteins encoded by RNA 2 allow virion formation and nematode-mediated transmission between plants (reviewed by MacFarlane, J. Gen. Virol., 80 2799 – 2807,1999).

The downregulation system is composed of separate cDNA clones of TRV RNA 1 and RNA 2 under the control of cauliflower mosaic virus (CaMV) 35S promoters on the transferred T-DNA of plant binary transformation vectors.

The TRV RNA 1 construct (pBINTRA6) contains a full-length infectious cDNA clone in which the RNA polymerase ORF is interrupted by intron 3 of the Arabidopsis Col-0 nitrate reductase NIA1 gene (Wilkinson and Crawford, Mol. Gen. Genet., 239 289 – 297, 1993), necessary to prevent expression of a TRV-encoded protein that is toxic to *E. coli*. This vector has been given the internal reference number p3209.

The TRV RNA 2 construct (pTV00), contains a multiple cloning site (MCS), leaving only the 5' and 3' untranslated regions and the viral coat protein (Ratcliff *et al.*, Plant Cell, 11 1207 – 1215, 1999). This vector has the internal reference number p3930 and contains a GatewayTM cassette and the gene of interest to be tested. The genes as presented in SEQ ID NO 1 to 21 are each cloned in this vector.

cDNAs were amplified using Gateway compatible primers and the cDNAs were entered into Entry Clones by BP recombination reactions. Subsequently the entry clones comprising the gene according to any one of SEQ ID NO 1 to 21 were checked via Ban2 restriction digest. The genes of interest were then entered into destination vectors by LR recombination reactions and the destination vectors were checked via ECORV restriction digestions. These expression clones were electroporated into the Argobacterium strain GV3101 agro and the plasmid pBintra6 was electroporated into pMP90 agro.

Inoculation

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To inoculate plants, *Agrobacterium* cultures carrying pBINTRA6 (strain C58C1RifR containing pMP90 plasmid) and pTV00 (strain GV3101 containing pMP90 plasmid) were grown and mixed and infiltrated to the leaves of *Nicotiana benthamiana* as previously described (English et al., Plant J., 12 597 – 603, 1997). Briefly, virus infection was achieved by *Agrobacterium*-mediated transient gene expression. *Agrobacterium* containing the TRV cloning vectors were grown overnight in L brith (+Tc+Km), *Agrobacterium* containing the helper plasmid was grown overnight in 10 ml YEB+Rif+Km. The culture was centrifuged and resuspended in 10 ml of 10mM MgCl₂, 1mM MES-pH5.6 and 100µM acetosyringone and kept at room temperature for 2 h. Separate cultures containing pBINTRA6 and TRV cloning vectors were mixed in a ratio of 1:10. The culture was then infiltrated to the underside of two leaves of three-weeks old plants using a 2 ml syringe without a needle. In two independent experiments 6 plants per agroabcterium clone were infected. In this way the cloned genes (SEQ ID NO 1-21) were transferred into the cells of the infiltrated region, and could be transcribed into the viral cDNAs in the leave cells. These transcripts then serve as an inoculum to initiate systemic infection of the plant. Consequently the VIGS system is activated, resulting in the downregulation of the

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host cell gene, corresponding to the cloned gene of interest. All experiments involving virus-infected material was carried out in controlled growth chambers. N. benthamiana plants were germinated ad grown individually on universal potting ground in pots at 25°C during the day (16h) and 20°C during the night (8h).

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The plants were phenotypically evaluated on a daily basis. Particular attention was given to visible leaf damage and growth inhibition. The effects of the suppression of gene activity using the VIGS system is measured by the phenotypic aspect of the plants, including leaf defects such as growth retardation, yellow or necrotic spots, early senescence, etc. The effects of the downregulation of genes identified by the methods of the invention are also measured on the flower structure and the flowering capacities of the transformed plants.

The severity of the phenotype is linked to the level of suppression of the geneactivity and indicates the degree in which the gene is essential for the plant Therefor the phenotype is an indication of the degree in which the gene is a valid target for a herbicide.

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Phenotypes of the infected plants.

1. Co-suppression of the gene leads to loss of gene transcription and protein expression in the virus infected leaf and induces leaf growth modification, including leaf wrinkling, curling, wilting, leading to cell death and/or plant death.

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2. Co-suppression of the geneleads to loss of gene transcription and protein expression in the virus infected leaf and induces leaf yellowing or senescence, or cell death and necrosis, leading to plant death.

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3. Co-suppression of the gene leads to loss of gene transcription and protein expression in the virus infected leaf and induces any of the following phenotypic symptoms: chlorotic regions around infection, crisp or crunchy leaf texture around infection, numerous surface lumps on either leaf surface, abnormal trichomes, abnormal leaf size, reduced growth, reduced final size, altered vascular leaf system, altered water movement in leaf, leading to cell death and/or plant death.

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4. Co-suppression of the gene leads to loss of gene transcription and protein expression in the virus infected leaf and induces any of the following anatomical symptoms: clumps of modified cells on the surface of the leaf (either abaxial or adaxial), individual cells detached from the epidermis, swollen or modified trichome cells, modification of leaf tissue structure, cell size, cell number, tissue composition, parenchyme, epidermis, etc, leading to cell death and/or plant death.

5. co-suppression of gene X leads to loss of gene transcription and protein expression in the virus infected leaf and induces any of the following biochemical symptoms, enzyme activity and products, degradation of leaf components and effects in neighboring leaves, stem, vascular system, degradation of cell wall structure, communication between cells, modification of cell-cell signaling leading to cell death and/or plant death.

The genes identified by the present invention can be utilized to examine herbicide tolerance mechanisms in a variety of plants cells, including gymnosperms, monocots and dicots. It is particularly useful in crop plant cells such as rice, corn, wheat, barley, rye, sugar beet, etc

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Example 3

Significant phenotypic alterations could be observed in plants infiltrated with Agrobacterium containing pBINTRA6 + Bstt44-4-340 (SEQ ID NO 18, acetolactate synthetase) and pBINTRA6 + Bstt2-42-520 (or T4-32-7) (SEQ ID NO 21, prohibitin) and pBINTRA6 + Bstt23-4-230 (SEQ ID NO 11, B-type CDK).

At 10days post-infiltration the first symptoms were visible. The symptoms were persistent until the end of the experiment and could be observed in at least 5 out of the 6 infiltrated plants.

The phenotypes of the plants transformed with acetolactate synthase are further described. In two separate replicated experiments, specific phenotypes on each plant infected with the 20 acetolactate synthetase downregulation construct were observed (Figure 2). Winkling and wrapping of the leaves as well as some chlorotic spots were observed. Thus acetolactate downregulation provoked a general growth arrest accompanied with chlorotic and necrotic areas. These observations were in line with previous reports, wherein acetolactate synthetase is described as a useful herbicide target.

The phenotypes of the plants transformed with prohibitin are further described.

In two separate replicated experiments, specific phenotypes on each plant infected with the prohibitin downregulation construct were observed (Figure 2). These plants showed strong wrinkling of the leaves about 20 days after infection, corresponding to the expected occurrence of silencing events. Thus the downregulation of probibitin provokes a severe leaf distortion and general growth arrest.

The phenotype of the plants inoculated with a B-type CDK downregulation construct are shown in Figure 3. A late (from 30 days after inoculation) but strong negative effect on the plant 35 growth was observed. The plants started to grow much slower and lost their apical dominance, resulting in the increased appearance of lateral branches.

Table 1. Functional classification of transcript tags

Function	Tags	S	G2	M	G1
		27.7%	15.8%	52.9%	3.6%
Cell cycle control	30	5/8 (0.078)	8/5 (0.068)	14/16 (0.114)	3/1
Cell wall	35	6/10 (0.047)	4/6 (0.136)	25/18 (7.1e⁻³)	0/1
Cytoskeleton	43	1/12 (1.2e ⁻⁵)	4/7 (0.090)	38/22 (2.1e⁻⁷)	0/2
Hormone response	13	6/4 (0.113)	1/2 (0.277)	6/7 (0.185)	0/0
Kinases/phosphatases ¹	27	4/8 (0.039)	1/4 (0.059)	19/14 (0.025)	3/1
Protein synthesis	50	15/14 (0.116)	5/8 (0.087)	29/26 (0.079)	1/2
Proteolysis	21	2/6 (0.026)	1/3 (0.144)	17/11 (0.039)	1/1
Replication and modification	74	57/20 (4.2e⁻¹⁹)	8/12 (1.0e ⁻⁵)	8/39 (1.0e ⁻¹⁸)	1/3
RNA processing	20	1/6 (6.8e-3)	1/3 (0.137)	18/11 (8.1e ⁴)	0/0
Signal transduction	10	1/3 (0.121)	3/2 (0.201)	6/5 (0.205)	0/0
Stress response	20	6/6 (0.192)	2/3 (0.229)	10/10 (0.159)	2/1
Transcription factors	27	4/8 (0.039)	10/4 (3.0e⁻³)	12/14 (0.112)	1/1
Transport and secretion ²	31	5/9 (0.047)	2/5 (0.076)	21/16 (0.031)	3/1
Unknown	175	37/48 (0.015)	19/28 (0.014)	112/93 (8.3e⁻⁴)	7/6

The total number of tags and the observed/expected number of tags within the different cell cycle phases for each functional group is given together with the probability values between parentheses as calculated based on the binomial distribution function, except for the G1-phase because the values were too small. A significant enrichment (P<e⁻³) of tags of a functional group within a particular cell cycle phase is indicated in bold.

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30 Table 2: overview of group 1 of sequences used for validation of candidate target genes

SEQ ID NO	CDS NO	Tag Name	Function	Fase
	2216	18R1850 C4-32-33 1E2	catalase	??
·		Bstt2-31-215	phytoene desasturase	??
3		Bstc13-1-145	L-ascorbate peroxidase	M-G1
<u> </u>		Bstc21-4-280	GTP-bindingprotein	M
7		Bstc33-2-310	vacuolarsortingreceptor	M
<u></u>		Bstc4-34-170	probable cinnamyl alcohol dehydrogenase	G1/S-S; M-G1
		Bstt34-3-470	kinesin	M
<u>/</u>		Bstt12-3-410	B-typeCDK	M
<u> </u>	2224	Bstt14-3-458	squalene mono-oxygenase	G1/S-S
10	2225	Bstt12-1-230	kinesin-likeprotein	M
11	2226	Bstt23-4-230	B-typeCDK	M
12	2227	Bstt2-42-225	B-typeCDK	M
13	2228	Bstt31-4-208	arabinogalactan protein precursor	G2/M-M
14	2229	Bstt 3-41-205	arabinogalactan protein precursor	G2/M-M

¹ Only kinases and phosphatases with unknown biological function.

² Except small GTP-binding proteins, which are classified under signal transduction.

WO 03/0	85115			
15	2230	Bstt33-4-285	late .	PCT/EP03/03703
16	2231	Bstt2-31-215	chorismate synthase kinesin-likeprotein	S-G2
17	2232	Bstt41-2-400	endo bata 4 4 4	M

1.0	2230	Bstt33-4-285	late - ·	1 C 1/EF03/03/03
16	2231	Bstt2-31-215	chorismate synthase	S-G2
17	2232	Bstt41-2-400	kinesin-likeprotein	M
18	2233	Bstt44-4-340	endo-beta-1,4glucanase	M
19	2234	G17-2-13 G17-2-13	acetolactate synthase	G2/S-G2-M-G1
20	2235	mapk9-ntf6.seg	WRKY transcription factor	20
21	2236	Bstt2-42-520	mapkinase phragmoplast associated N	TF6 22
		.2 020	prohibitin	22
able 2.		_		

Table 3: overview of group 2 sequences of full-length sequences that are cell cycle modulated and of which some are involved in the cell cycle process

1	EQC	JO	Gene name Gene name
ID N	INC	J	
22			Protein kinase mRNA, complete, N. tabacum, 2073 bp
23			BY2 AA041K03 probable DNA-binding protein GBP16 - rice T02069, N. tabacum, 834 bp
24	06	15	BY2 AA042C09 probable nuclear DNA-binding protein GBP16 - rice T02069, N. tabacum, 834 bp tabacum, 1185bp
	l		tabacum, 1185bp Tobable nuclear DNA-binding protein G2p [imported] in Arabidopsis T51151 N
25	061	16	BY2.4004417
26	061	7	BY2 A00417 transcription regulator-like in Arabidopsis AB025604 N. 4-1-
	100.		BY2-AA044J17 transcription regulator-like in Arabidopsis AB025604, N. tabacum, 1893bp AA044J23 ATP-dependent RNA helicase CA3 of the DEAD/DEAH box family; Dbp3p; BY2- BY2-AA046C45 BY2-AA046C45 BY2-AA046C45
27	061	-	COUNTY OF THE PROPERTY OF THE
8			DIETONOGO, IS DICTOR PROCESS OF THE CONTROL OF THE
	061	9	BY2-AA047G13 14-3-3-like protoin C Poole II Alabidopsis BABU8417 AB025622 N tabacum 7225
9	062		DIET VIVIUM IN INCOME A CONTRACTOR OF THE PROPERTY OF THE PROP
0	062	1	BY2-AA066H11P19H05 phosphoprotein phosphatase 2A regulatory chain T03684 N. tabacum, 1764 t
1	062	2	BY2-AA0601 10 tropping prosphoprotein phosphatase 2A regulatory chain T03694 N Anh
2	062	3	BY2-AA069L10 transcription factor-like protein in Arabidopsis BAB09482 AB012246, N. tabacum, 1764 BY2-AA073K06 SET protein, phospatase 2A inhibitor in Arabidopsis AAC52377 4-AO04000 abacum, 831
	102.	4	b 12-AA073K06 SET protein, phospatase 2A inhibitor in Archidentic Academy 831
3	0624		
<u>3</u> 4		. ,,	215-00U(3WP19EU/phoophe
+	0625) [E	BY2-AA073MP19B07 phosphoprotein phosphatase 2A regulatory chain T03684, N. tabacum, 1764bp 83bp BY2-AA076003B40B09 Market Communication
_	 		
5	0626		
3_	0627	TE	BY2-AA079J13 putative casein kinase I in Arabidopsis AAG51841.1 AC010926_4, N. tabacum, 2514 bp BY2-AA080G14 porin I 36K in potato S46959, N. tabacum, 393bp
7	0628	F	3Y2-AA080C14 paris Policius Casen kinase I in Arabidopsis AAG51841 1 AC010036
	0629	_ , -	7 - 7 17 17 17 17 17 17 17 17 17 17 17 17 1
	0020		172- AA081P13p21E02 separation anxiety protein like in Assistantia
1	0000	- <u> a</u>	3Y2- AA081P13p21E02 separation anxiety protein-like in Arabidopsis CAB96669.1 AL360314, N.
_	0630	10	William Sty Copy of Occo. N
	0631	В	Y2-AA085N17p21H04 14-3-3-like protein in potato 16R P93784 N. tabacum 768bp
	0632	B	Y2-AA087C16p21G03 AP2 domain transcription factor homolog in potato T07784 N. tabacum, 891bp
	0633	B	Y2-A088B13 Process AP2 domain transcription factor homolog in potato T07784 N.A.
- 1		12	48bp Putative RING zinc finger protein in Arabidopsis CAB80936.1 At 161491 N. tabacum, 891bp
	0634		
	0635		
۲	0000	I B	Y2-AA096M07 peptidyl-prolyl cis-trans isomerase-like protein BAB10691.1 AB015468 N. tabacum
		450	Obp A Data Somerase-like protein BAB10691 1 AB015469 N. L.
_(636		
_ 0	637	BY	2-AA096M22 cell division-like protein in Arabidopsis BAB09106.1 AB017069 N. tabacum 1518bp (2-AA098B08p21D11 similarity to DAG protein in Arabidopsis T45963 N. tabacum 687bp
0	638	1 RY	2-AAOGRAGO AND
-	_	1114	12 Nosebbook 2 10 11 similarity to DAG protein in Arabidonsis BAAA2200 4 100
1	638	2 12	22-AA098B08p21D11 similarity to DAG protein in Arabidopsis T45963 N. tabacum 687bp AA091G15-24505 N. tabacum 087bp
ال	639	RA5	2-AA109N15 GAMM1 protein-like in Arabidopsis BAB08430.1 AB017067 N. tabacum 888bp, (MYG1)
+		FAN	AILY, proliferation associated AND
	640		
100	341	BY2	AAA114N16 unknown protein in Arabidopsis BAB03019.1 AP001297; candidate tumor suppressor AAA114N21p231p2320204458p
DΕ	342	EVO	ING1 homolog in Homo sapiens N. tabacum 720bp
_	343		
٥	/4 0	BYZ	C-AA119N11p22G04 serine/threonine-specific protein Ar201456 1N. tabacum 699bp
<u>_</u>		nava	CUID 1793DD
	62		
06	63	BY2	-AA043A01 >qb AA024540 4054555 protein T17F15.80 - Arabidopsis thaliana
	li li	HVICC	Mana tahacumi
06	64	RV2	AAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA
	- · · [- 1 Z-	AMU44CUZ > GDJJBAAO2028.1] (D11470) chloroplast clareating in T
16	<u> </u>	anac	AA044C02 >dbj BAA02028.1 (D11470) chloroplast elongation factor TuB(EF-TuB) [Nicotiana
J6(36 E	3Y2-	AA045P04p01G10 spi(043681)NUTD 10154) gene id:MYN8.3~pir T02891~similar to unknown protein
		22.40	AA046C08p19E02 dbj BAB30364.1 (AK016659) putative [Mus musculus]
966	o/ ⊩	3 Y フェル	AADARC TIRROTOEDO AL TOA DE COMENTA DE COMEN

	WO 0	3/085115 PC1/EP03/03/03
60	0668	BY2-AA046E06 pir T50556 stamina pistilloidia protein Stp [imported] - garden pea
		BY2-AA046G14 dbj BAB26082.1 (AK009117) putative [Mus musculus]
	0670	BY2-AA046H23 emb CAA98172.1 (Z73944) RAB8A [Lotus japonicus]
	0671	BY2AA048A05 gb]AAD15504.1] (AC006439) putativeAAA-type ATPase [Arabidopsis thaliana]
	0672	BY2-AA049K03 dbj BAB24909.1 (AK007240) putative [Mus musculus]
	0673	BY2-AA051A10 dbj[BAB02543.1] (AP000417) mitotic checkpoint protein [Arabidopsis thaliana]
66	0674	BY2-AA051L22p19H03 gb AAD48948.1 AF147262_11 (AF147262) contains similarity to Pfam family
		PF00400 -WD domain
67	0675	BY2-AA052E10 >gb AAF52905.1 (AE003628) CG4968 gene product [Drosophila melanogaster]
68	0676	BY2-AA052F14 >gb AAF79819.1 AC007396_20 (AC007396) T4O12.22 [Arabidopsis thaliana]
	0677	BY2-AA052G16p19D04 >dbj BAB09843.1 (AB005246) gene_id:MUP24.12~unknown protein
		[Arabidopsis thaliana]
70	0678	BY2-AA052N17 >gb AAG42914.1 AF327533_1 (AF327533) unknown protein [Arabidopsis thaliana]
	0679 1	BY2-AA053C11.1 >dbj BAB22857.1 (AK003561) putative [Mus musculus]
	0670	2 BY2-AA053C11.2 >gb[AAC62883.1] (AC005397) hypothetical protein [Arabidopsis thaliana]
	0679_	2 BT2-AA035CT1.2 - gulAAC026G3.1 (AC003537) hypothetical protein prisonal prisonal protein prisonal prisonal protein prisonal protein prisonal prisonal protein prisonal
73		BY2-AA062A09 >gb AAF01061.1 AF189284_1 (AF189284) nucleolar G-protein NOG1 [Trypanosoma
		brucei)
	0681	BY2-AA062G03 >pir T02135 hypothetical protein F8K4.10 - Arabidopsis thaliana
75	0682	BY2-AA065E08 >pir T00795 hypothetical protein F24L7.13 - Arabidopsis thaliana
76	0683	BY2-AA072K18 >emb CAB40381.1 (AJ010819) GrpE protein [Arabidopsis thaliana]
77	0684	BY2-AA075K12 >gb AAD31331.1 AC007354_4 (AC007354) T16B5.4 [Arabidopsis thaliana]
	0685	BY2-AA076N08 >dbi BAA94770.1 (AP001859) ESTs AU082761(S5084) D42006
	0686	BY2-AA080D01 >gb AAF80646.1 AC012190_2 (AC012190) Contains similarity to F28O16.19 a putative
1, 3	0000	translation initiation protein
00	0607	BY2-AA081P14 >gb AAD32777.1 AC007661_14 (AC007661) unknown protein [Arabidopsis thaliana
	0687	BY2-AA082H04p21F02 >dbj BAB10171.1 (AB016880) gene_id:MTG10.12~pir T05795~strong similarity
81	0688	
L		to unknown
	0689	BY2-AA082H06p21G04 >pir T09039 hypothetical protein F26K10.110 - Arabidopsis thaliana
83	0690	BY2-AA082M07p21B05 >dbj BAB01783.1 (AB022215) gene_id:MCB17.19~unknown protein
]		[Arabidopsis thaliana]
84	0691	BY2-AA083B24p21C04 >dbj BAB08247.1 (AB006698) gene_id:MCL19.6~unknown protein [Arabidopsis]
1	ł	thanliana)
85	0692	BY2-AA083C05p21D02 >gb AAH02924.1 AAH02924 (BC002924) Unknown (protein for
۲	0002	IMAGE:3956179) [Homo sapiens]
86	0693	BY2-AA085D08p21C05 >pir T47624 hypothetical protein T5N23.10 - Arabidopsis thaliana
_	0694	BY2-AA085F09p21H01 >gb[AAF79503.1 AC002328_11 (AC002328) F20N2.15 [Arabidopsis thaliana]
		BY2-AA085M15p21D04 >gb AAF97305.1 AC007843_8 (AC007843) Unknown protein [Arabidopsis
88	0695	
-		thaliana)
89	0696	BY2-AA088K23p21G05 >gb AAG52001.1 AC012563_11 (AC012563) unknown protein; 64612-65506
L	<u> </u>	[Arabidopsis thaliana]
90	0697	BY2-AA088L24p21A07 >gb AAD55292.1 AC008263_23 (AC008263) Contains PF 00249 Myb-like DNA-
j	ļ	binding domain.
91	0698	BY2-AA089F12p21H05 >gb AAD55274.1 AC008263_5 (AC008263) Strong similarity to gb D21805
	1	calcium-dependent protein kinase
92	0699	BY2-AA089M17 >pir T02186 hypothetical protein F14M4.16 - Arabidopsis thaliana
		BY2-AA090J23p21G08 >pir T48545 hypothetical protein F14F18.30 – Arabidopsis thaliana
	0700	BY2-AA090323p21G08 > pirj[146345 hypothetical protein 1141 16.50 = Arabidopsis indicates BY2-AA092F12p21H06 > emb CAB46854.1 (AJ388555) hypothetical protein [Canis familiaris]
	0701	D 12-A-MUSZC 12/2 [100 Zenia] AD 4004-01-1 [AUS00000] Hypothetical protein [Conia familians]
95	0702	BY2-AA092L20p21E07 >gb[AAD10646.1] (AC005223) 45643 [Arabidopsis thaliana]
96	0703	BY2-AA093J23p21C11 >gb AAG51461.1 AC069160_7 (AC069160) unknown protein [Arabidopsis
	<u> </u>	thaliana)
97	0704	BY2-AA093L18p21D09 > emb CAC15504.1 (AJ297917) B2-type cyclin dependent kinase [Lycopersicon
98	0705	BY2-AA093M19 >gb AAG12535.1 AC015446_16 (AC015446) Unknown protein [Arabidopsis thaliana]
	0706	BY2-AA094B12p21F10 >dbj BAB02118.1 (AP000381) contains similarity to unknown
100	0707	BY2-AA096G05p21A11 dbj BAB02118.1 (AP000381) contains similarity to unknown
		CICI AA094B12p21F10
		BY2-AA097G22p21D10 >gb AAG60065.1 AF337913_1 (AF337913) unknown protein [Arabidopsis
1102	0708	
100	l	thaliana
103	0709	BY2-AA099F04 gb AAG52457.1 AC010852_14 (AC010852) hypothetical protein; 12785-11538
		[Arabidopsis thaliana]
104	0710	BY2-AA099N08p21H09 gb AAK14411.1 AC087851_3 (AC087851) unknown protein [Oryza sativa]
	0711	Ict AA100B09 reftNP_009820.1I Ybr261cp [Saccharomyces cerevisiae]
	0712	BY2-AA109N02 ref[NP_002848.1] peroxisomal farnesylated protein; Housekeeping gene 33kD [Homo
	[sapiens
107	0713	BY2-AA114E09p22F02 pir T51434 hypothetical protein F2G14_10 - Arabidopsis thaliana
	0714	BY2-AA115B14p22C02 dbj[BAB08888.1] (AB012243) gene_id:MIJ24.6~ref[NP_013897.1~similar to
100	D/ 14	
L	1	lunknown protein

WO 03/085115
109 10715 18Y2-AA115E08p22C04 SchiDVO AA1199999 44119

109	0715	BY2-AA115F08p22C04 SekiDV0 AA1100000 W W
	ĺ	BY2-AA115F08p22C04 >gb BY2-AAH03900.1 AAH03900 (BC003900) Similar to hypothetical protein
110	0716	DV3 Additional Processing
' ' '	0,10	BY2-AA115L12p22G01 >gb AAF43925.1 AC012188_2 (AC012188) Contains similarity to PIT1 from Arabidopsis thaliana
444	0747	Arabidopsis thaliana Garage Account 188 (ACO12188) Contains similarity to PIT1 from
1111	0717	BY2-AA116L23p22E01 >dbj BAB01460.1 (AP000731) gene_id:MCB17.21~unknown protein
<u> </u>		[Arabidopsis thaliana]
1112	0718	BY2-AA117B12p21G12 >sp O23708 PSA2_ARATH PROTEASOME SUBUNIT ALPHA TYPE 2 (20S
<u> </u>		PROTEASOME ALPHA SUBUNIT B)
113	0719	BY2-AA117E08p32A02 > pil/E04405
114	0720	BY2-AA117E08p22A03 >pir F81195 conserved hypothetical protein NMB0465 [imported] - Neisseria
	0721	BY2-AA117O08p22E03 >dbj BAB01753.1 (AP000603) gb BY2-AAD10646.1~gene_id:MRP15.12
	0722	BY2-AA118D23p22E02 >emb CAB89490.1 (AJ277062) CRK1 protein [Beta vulgaris], cdc2 like kinase
1110	0/22	BY2-AA119D12p22H04 >dbj BAB01163.1 (AP000410) gene_id:K10D20.9~unknown protein
		[Arabidopsis thaliana]
	0723	BY2-AA120G12 >gb/BY2-AAB63649 11 (AC001645)
118	0724	BY2-AA120G12 >gb BY2-AAB63649.1 (AC001645) hypothetical protein [Arabidopsis thaliana] BY2-AA120G19p22D05 >gb BY2-AAF69547.1 AC008007_22 (AC008007) F12M16.18 [Arabidopsis thaliana]
1	ļ	thaliana1 thaliana1 thaliana1 thaliana1 thaliana1 thaliana1
		in the state of th

Table 4: overview of group 3 sequences that show homology with proteins of unknown function

0.0	OTer		7			
ID.	Q Tag name a	nd	Function		Fase	_
NO	1				1	
11			 		ļ	
12		_			M-G1	_
12			 		G2/M-M-G1	
12			<u> </u>		G2/M-M-G1	
12:	3 Bstc1-13-143		unknownprotein	_	G2/M-M-G1	
124	4 Bstc11-3-190		unknownprotein		G2/M-M-G1	
12					M-G1	
126			putativeprotein		G2/M-M-G1	
127			·		G1/S; M-G1	
128			unknown		M-G1	_
129			hypotheticalproteir	1	S-G2	
130		_	hypotheticalproteir		M-G1	_
131		-			G2/S-G2	
132		-			G2/S-G2	
133			unknown		M-G1	
134			unknown	1	G2/M-M-G1	
135		!	hypotheticalprotein		G2/M-M	_
136		_/	nypotheticalprotein		G2/M-M-G1	7
137			unknown		VI-G1	٦
138		1	unknownprotein	(G2-M-G1	7
	Bstc2-22-100	- 1	ınknown	1	32-G2/M	7
139		1		10	32-M	1
140	Bstc2-22-240	_	nypotheticalprotein	_:	Λ	1
_	Bstc22-2-270	1		C	91/S; M-G1	1
142	Bstc2-23-135	4		C	62/S-G2-M	1
143	Bstc2-23-220		ınknown	C	62-M-G1	1
144	Bstc22-4-215	<u>lh</u>	ypotheticalprotein		62/M-M	1
145	Bstc2-31-280	1		G	62/M-M-G1	1
146	Bstc23-2-240		nknown	N	1	1
147	Bstc23-2-330	р	utativeprotein	N	1	
148	Bstc23-2-370	1		G	1/S-S;	
149	D-1 0 00	1		G	2/M-M-G1	
149	Bstc2-32-400			G	1/S-S;	i
150	D-1 00 0	1		G	2/M-M-G1	
	Bstc23-3-270	\downarrow		G	1/S-S; M-G1	
	Bstc2-33-280	u	nknownprotein	O	1/S-S;M-G1	
	Bstc2-34-120		nknown	G	2/M-M-G1	
153 154	Bstc23-4-300	ur		M		
	Bstc2-41-165	_		G	1/S-S	
155	Bstc2-42-100	ur	nknown	G	1/S-S	
156	Bstc2-43-210				·G1	
	Bstc31-185	ur	known	G	2/M-M-G1	
158	Bstc3-12-145	ur	known ·	S-	G2	
159	Bstc3-12-290	ur			2/M-M-G1	

ID D	i	nd	Function	Fase
16			l um lem a com	
16			unknown	G2/M-M-G1
16	2 Bstc3-21-125		unknown	M-G1
10.	2 DSIC3-21-125)	l	G1/S-S;
16:	2 Pata 20 0 4 5 0			G2/M-M-G1
110	3 Bstc32-2-150)	putativeprotein	G1/S-S;
16	1 0-4-00 4 400			G2/M-M-G1
164 165		-		
1100	5 Bstc32-4-370	١.		G1/S-S-G2/S
100	2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2			M-G1
166 167			putativeprotein	G1/S-S-G2/S
167	Bstc33-2-145	- 1	hypotheticalproteir	G1/S-S;
400				G2/M-M-G1
168				G1/S-S
169			outativeprotein	G2/M-M-G1
170			unknown	G2/M-M
171		l	unknown	M-G1
172				G2/M-M-G1
<u> 173</u>	Bstc3-41-360	7		G2/M-M-G1
74		\exists		M-G1
75	Bstc3-43-135	7	· · · · · · · · · · · · · · · · · · ·	G1
76	Bstc3-43-180	\top		M-G1
77	Bstc3-43-193	U	ınknown	G1/S-S;
				G2/M-M-G1
78	Bstc3-43-287	\top	·	G1/S-S
79	Bstc3-44-145	+		M-G1
80	Bstc3-44-375	'n	utativeprotein	M-G1
81	Bstc4-11-120	- P	ypotheticalprotein	
82	Bstc4-11-320		nknown	G2/M-M-G1
83	Bstc42-3-115	_		M-G1
<u>84</u>	Bstc42-3-125		nknown	M-G1
B5	Bstc4-23-210	P	utativeprotein	G2/M-M-G1
B6	Bstc42-4-225	-	-1	M-G1
37	Bstc4-32-115		nknown	G1/S-S-G2
,	108104-32-115	Jui	nknownprotein	G1/S-S;
38	Poto4 22 405	1		G2/M-M-G1
39	Bstc4-32-185	_	nknown	G1/S-S
	Bstc4-32-190	_		G2/M-M
	Bstc4-32-270	ur		G2/S-G2-M
71	Bstc4-32-410			G1/S-S-G2-
اَـــا		_		G2/M
_	Bstc4-34-250			G2/M-M-G1
_	Bstc4-41-230	þг		G2/M-M-G1
	Bstc4-43-113	un		M-G1
5	Bstc44-3-125	Γ		G2/M-M

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	WO 03/0851	115	
196	Bstt1-12-340	unknown	G2/M-M
197	Bstt12-2-225		G1/S-S-G2
198	Bstt1-22-330	unknown	G2/M-M-G1
199	Bstt12-2-420	unknownprotein	G2/M-M-G1
200	Bstt12-2-540	hypotheticalprotein	G2/M-M-G1
201	Bstt1-23-155		M-G1
202	Bstt12-3-215	hypotheticalprotein	G2/M-M-G1
203	Bstt12-3-280	unknown	G1/S-S-G2
204	Bstt12-3-310	hypotheticalprotein	G1/S-S
205	Bstt12-3-350		G1/S-S-G2-
Ĺ			G2/M
206	Bstt1-24-205		G2/M-M-G1
207	Bstt1-24-220		G1/S-S-G2
208	Bstt1-31-170	hypotheticalprotein	G2/M-M-G1
209	Bstt1-31-215	unknown	G2/M-M-G1
210		unknown	G2/M-M-G1
211	Bstt14-4-310	unknownprotein	G2/M-M-G1
212	Bstt2-11-165	unknown	G2/M-M-G1
213	Bstt2-12-190		G1/S-S-G2
214	Bstt21-4-150	hypotheticalprotein	G1/S-S-G2/S
215	Bstt21-4-250		G1/S-S;
			G2/M-G1
216	Bstt21-4-470	<u> </u>	G2/M-M-G1
217	Bstt22-1-170	unknown	S-G2
218	Bstt2-21-190	unknown	G2/M-M
219	Bstt22-2-190	unknown	G2/M-M-G1
220	Bstt22-2-290	hypotheticalprotein	G2/M-M-G1
221	Bstt22-3-225		M
222	Bstt22-3-275	unknown	G2/M-M
223	Bstt22-3-315	TomatoEST	G2/M-M-G1
224	Bstt22-3-370	unknown	G2/M-M-G1
225	Bstt22-3-390	putativeprotein	G2/M-M-G1
226	Bstt22-3-480		G2/M-M-G1
227	Bstt23-1-140		S-G2-G2/M
228	Bstt23-120	unknownprotein	G2/M-M-G1
229	Bstt23-1-200		S-G2-M
230	Bstt2-31-300	unknown	S
231	Bstt2-32-220	<u> </u>	M
232	Bstt2-32-400	hypotheticalprotein	G2/M-M-G1
233	Bstt23-3-350	unknown	G2-M
234	Bstt23-370	unknown	G2/M-M-G1
235	Bstt24-1-320	 	S-G2
236	Bstt24-2-310	 	G2/M-M-G1
237	Bstt2-43-210	unknown	G2-M
238	Bstt2-43-240		S-G2/S
239	Bstt31-1-100	hypotheticalprotein	G1/S-S-G2

	WO 03/0851	19				1 C 1/E	PUS/US/US
	Bstt1-12-340	unknown	G2/M-M	240	Bstt3-11-205		G1/S-S-G2
197	Bstt12-2-225		G1/S-S-G2	241	Bstt31-1-250	hypotheticalprotein	G2/M-M-G1
	Bstt1-22-330	unknown	G2/M-M-G1	242	Bstt31-1-430	hypotheticalprotein	G2/M-M-G1
	Bstt12-2-420	unknownprotein	G2/M-M-G1	243	Bstt3-12-360	unknownprotein	G2/M-M
200	Bstt12-2-540	hypotheticalprotein	G2/M-M-G1	244	Bstt31-3-380		G1/S-S
201	Bstt1-23-155		M-G1	245	Bstt31-4-420	hypotheticalprotein	G2/M-M-G1
202	Bstt12-3-215	hypotheticalprotein	G2/M-M-G1	246	Bstt32-180	putativeprotein	G2-M-G1
203	Bstt12-3-280	unknown	G1/S-S-G2	247	Bstt3-22-160	PotatoEST/hypothel	G1/S-S-G2
204	Bstt12-3-310	hypotheticalprotein	G1/S-S	-		icalprotein	_
205	Bstt12-3-350		G1/S-S-G2-	248	Bstt32-3-175	unknown	G2/M-M
			G2/M	249	Bstt32-3-325	unknown protein	G2/M-M-G1
206	Bstt1-24-205		G2/M-M-G1	250	Bstt3-24-135	unknown	G2/M-M-G1
207	Bstt1-24-220		G1/S-S-G2	251	Bstt3-24-200	·	G2/M-M-G1
208	Bstt1-31-170	hypotheticalprotein	G2/M-M-G1	252	Bstt3-31-215	unknownprotein	G2/M-M-G1
209	Bstt1-31-215	unknown	G2/M-M-G1	253	Bstt3-31-330	unknown	G1/S-S-G2
	Bstt13-210	unknown	G2/M-M-G1	254	Bstt33-1-350	unknown	G2/M-M-G1
211	Bstt14-4-310	unknownprotein	G2/M-M-G1	255	Bstt33-1-510	putativeprotein	G2/M-M-G1
212	Bstt2-11-165	unknown	G2/M-M-G1	256	Bstt33-3-220	unknown	G2/M-M-G1
213	Bstt2-12-190		G1/S-S-G2	257	Bstt33-3-245	unknownprotein	G2/M-M-G1
214	Bstt21-4-150	hypotheticalprotein	G1/S-S-G2/S	258	Bstt3-33-550	hypotheticalprotein	G1/S-S; M-G1
215	Bstt21-4-250		G1/S-S;	259	Bstt33-4-140	putativeprotein	S-G2
L			G2/M-G1	260	Bstt34-2-165	unknown	G1/S-S-G2
216	Bstt21-4-470		G2/M-M-G1	261	Bstt3-42-325	hypotheticalprotein	G2/M-M-G1
217	Bstt22-1-170	unknown	S-G2	262	Bstt3-44-150	unknown	G2/M-M-G1
218	Bstt2-21-190	unknown	G2/M-M	263	Bstt3-44-250	unknown	G2/M-M-G1
219	Bstt22-2-190	unknown	G2/M-M-G1	264	Bstt34-4-310	unknown	G2/M-M-G1
	Bstt22-2-290	hypotheticalprotein	G2/M-M-G1	265	Bstt3-44-345	hypotheticalprotein	G2/M-M-G1
	Bstt22-3-225		M	266	Bstt41-2-340		G2/M-M-G1
222	Bstt22-3-275	unknown	G2/M-M	267	Bstt41-3-310	unknown	G2/M-M
	Bstt22-3-315	TomatoEST	G2/M-M-G1	268	Bstt4-21-185		M-G1
	Bstt22-3-370	unknown	G2/M-M-G1	269	Bstt42-1-370		S-G2-G2/M
225	Bstt22-3-390	putativeprotein	G2/M-M-G1	270	Bstt4-23-480	unknown	G2/M-M-G1
226	Bstt22-3-480		G2/M-M-G1	271	Bstt4-24-170		G2/M-M-G1
	Bstt23-1-140		S-G2-G2/M	272	Bstt43-265	unknown	G1/S-S-G2/M
	Bstt23-120	unknownprotein	G2/M-M-G1	273	Bstt43-3-350	unknown	G2/M-M-G1
	Bstt23-1-200		S-G2-M	274	Bstt4-33-390	hypotheticalprotein	G1/S-S;G2/M-
	Bstt2-31-300	unknown	S				M-G1
231	Bstt2-32-220		M	275	Bstt4-34-280		G2/M-M-G1
	Bstt2-32-400	hypotheticalprotein	G2/M-M-G1	276	Bstt43-4-300	unknownprotein	G2/M-M-G1
	Bstt23-3-350	unknown	G2-M	277	Bstt43-4-330	unknownprotein	G2/M-M-G1
	Bstt23-370	unknown	G2/M-M-G1	278	Bstt43-4-340		G2/M-M-G1
	Bstt24-1-320		S-G2	279	Bstt44-4-250	hypotheticalprotein	G2/M-M
	Bstt24-2-310		G2/M-M-G1	280	Bstt4-44-400	hypotheticalprotein	G2/M-M-G1
	Bstt2-43-210	unknown	G2-M	281	MBc03-90	unknown	S-G2
238	Bstt2-43-240		S-G2/S	282	MBc42-180	unknown	G2-M-G1
239	Bstt31-1-100	hypotheticalprotein	G1/S-S-G2	283	MBc43-210	unknown	G1/S-S-G2

Table 5: overview group 4 sequences showing no homology to known genes

SEQ	Tag name	Function	Fase
ID			1
NO		İ	
284 (Bstc1 1-100	unknown	G2/S-G2-M
285 I	Bstc1 -11-110	unknown	S
286	3stc1 -11-115	unknown	G1/S-S;G2/M-M-G1
287 [Bstc1 -11-120		G1/S-S-G2
288	Bstc1 1-1-125	unknown	G2/M-M-G1
289	Bstc1 1-1-290	NaD	G1/S;G2/M-M-G1
290	3stc1 -12-155		G2/S-G2-M
291	8stc1 -12-175	unknown	S
292	Bstc1 -12-185	unknown	G2/M-M-G1
293	Bstc1 1-3-116	unknown	S-G2
294	3stc1 1-3-118	unknown	G2/M-M-G1
	Bstc1 -13-120		S
296	3stc1 -13-130	Í	G1/S-S; G2/M-M-G1
297	Bstc1 -13-132	unknown	M-G1

ng no homology to known genes					
1	Function	Fase			
ID	[ĺ			
NO					
298 Bstc1 -13-142	unknown	G1/S-S			
299 Bstc 11-3-187	unknown	S-G2/S			
300 Bstc1 1-3-200	unknown	G1/S-S-G2/S			
301 Bstc1 1-3-290	unknown	G2/S-G2-M-G1			
302 Bstc1 -14-100	unknown	G2/M-M			
303 Bstc1 -14-108	unknown	G2/M-M-G1			
304 Bstc1 1-4-130	unknown	G1/S-S-G2			
305 Bstc1 1-4-135	unknown	G2/M-M-G1			
306 Bstc1 1-4-140	unknown	S-G2-M			
307 Bstc1 -14-155		G2/M-M			
308 Bstc1 -14-165		G2-G2/M			
309 Bstc1 -14-167		G2-G2/M			
310 Bstc1 1-4-175		G2/M-M-G1			
311 Bstc1 1-4-200	unknown	G1/S-S			

WO 03/085115

312 Bstc1 2-1-110	unknow	n	<u> S</u> -G2
313 Bstc1 -21-150	unknow	n	G2/M-M-G1
314 Bstc1 2-1-160			G2-M-G1
315 Bstc1 2-1-240	unknow		
316 Bstc1 2-1-95	unknow		M-G1
317 Bstc1 -22-110	GIRTOW	<u> </u>	G1/S-S-G2
318 Bstc1 2-3-103			G2-M-G1
319 Bstc1 2-3-125			G2/M-M-G1
320 Bstc1 2-3-235			G1/S-S; G1
321 Bstc1 2-3-237			M-G1
322 Bstc1 2-4-130			G1/S-S
323 Bstc1 2-4-133			G2/M-M - G1
324 Bstc1 2-4-145	unknown		S-G2
325 Bstc1 2-4-235	unknown		VI-G1
326 Pote 1 2 4 4 50	unknown		32/M-M-G1
326 Bstc1 3-1-150			Л-G1
327 Bstc1 3-2-170	unknown		32/M-M-G1
328 Bstc1 3-2-180	unknown		91/S-S
329 Bstc1 3-2-190	unknown	C	S1/S-S
330 Bstc1 3-2-280	unknown		61/S-S; G2/M-M-G1
331 Bstc1 -41-170	unknown	G	61/S-S
332 Bstc1 -41-175	unknown		61/S-S
333 Bstc1 -41-180	unknown		1/S-S; G2/M-M-G1
334 Bstc1 -41-210	unknown		1/S-S
335 Bstc1 -41-230			1/S; G2/M-M-G1
336 Bstc1 4-2-140	unknown	- 10	-G1
337 Bstc1 -42-150	unknown		
338 Bstc1 -42-80	unknown		2/S-G2
339 Bstc1 -42-90	unknown		1/S-S-G2
340 Bstc1 -43-105	UTKTOWN		2-M
341 Bstc1 4-3-105			2/M-M
342 Bstc1 -43-110		G	1/S-S; G2/M-M
0.40		G.	1/S-S; G2-M
10	unknown	G2	2/M-M-G1
345 Poted 43-140	unknown		G2
345 Bstc1 -43-150		G2	2/M-M-G1
346 Bstc1 -43-175			G2
347 Bstc1 -43-185	unknown	G1	/S-S-G2/S
348 Bstc1 4-3-235	unknown		/S-S
349 Bstc1 4-3-260	unknown	G2	/M-M-G1
350 Bstc1 -43-65	unknown	G1	/S-S-G2
351 Bstc1 -43-75	ınknown	S-C	
352 Bstc1 -44-138	ınknown		/S-S-G2/S
353 Bstc1 -44-140 L	ınknown	G2	/S-G2-M
354 Bstc1 -44-157	inknown	G2/	'S-G2
355 Bstc1 4-95	inknown	G2/	M-M
356 Bstc2 1-1-100 lu	nknown	G2/	M-M-G1
357 Bstc2 1-1-140 lu	nknown		S-S-G2
358 Bstc2 1-1-145 u	nknown	M-C	
359 Bstc2 1-1-65 u	nknown		M-G1
360 Bstc2 1-2-120			M-M
361 Bstc2 1-2-215			
362 Bstc2 1-2-75		02/	M-M
363 Bstc2 -13-110		S-G	
	aknow:-	01/	S-S;G2/M-M
000	nknown		M-M-G1
13.00	nknown	M-G	
1267 6 . 6	rknown		Л-M-G1
368 Beto2 14-130 Ur	known	G2/N	Л-M-G1
368 Bstc2 -14-135 ur	known	S-G2	
369 Bstc2 1-4-135		S-G2	
370 Bstc2 1-4-155 un	known	G2/N	1-M-G1
371 Bstc2 -14-160		M-G	1
	known	G2/S	
3/3 Bstc2 2-100 lun		G2-N	
374 Bstc2 -21-120 lun		G1/S	
375 Bstc2 2-1-125 Jun		S-G2	
		M-G1	

		PCT/EP03/03703
377 ₿ stc22-1-98	unknov	•
378 Bstc2 2-2-11		
379 Bstc2 -22-16		
380 Bstc2 2-2-16	5 unknow	
381 Bstc2 -22-90		S; G2-M
382	unknow	n G2/M-M
383 Bstc2 -23-14	0	M-G1
384 Bstc2 2-3-150	5	S-G2
385 B stc2-23-17	5	M-G1
386 Bstc2 -23-195	unknow	
387 Bstc2 2-3-90		M-G1
388 Bstc2 -24-100		
389 Bstc2 2-4-140		G1/S-S-G2-M
390 Bstc2 -24-165		G2/M-M
391 Bstc2 -24-170		1 G1/S-S
392 Bstc2 -31-140	unknowr	G2/M-M-G1
393 Bstc2 -31-160		M-G1
394 Bstc2 -31-170	unknowr	
000 PSICE 3-2-139	unknowr	G2/M-M-G1
396 Bstc2 -32-285		G2/M-M
397 Bstc2 3-2-360	unknown	
	unknown	
399 Bstc2 3-3-175	unknown	
400 Bstc2 -33-200	unknown	G2/M-M-G1
401 Bs tc23-3-305	unknown	M-G1
402 Bstc2 -33-85	<u> </u>	S-G2
403 Bstc2 -33-95	unknown	G2/M-M-G1
404 Bstc2 3-4-110	unknown	G2-M
405 Bstc2 3-4-120	unknown	G1/S-S-G2
406 Bstc2 3-4-310		S-G2
407 Bstc2 3-4-335		G2-M-G1
408 Bstc2 -41-110	unknown	S-G2
409 Bstc2 4-2-165	-	M-G1
410 Bstc2 -43-105	unknown	S-G2-G2/M
411 Bstc2 -43-130	unknown	G2/M-M
412 Bstc2 4-3-285		G1
413 Bstc2 -43-77	unknown	G2/M-M-G1
414 Bstc2 -43-90	unknown	G2/M-M-G1
415 Bstc2 4-4-125	unknown	G1/S-S
416 Bstc2 -44-175 417 Bstc2 4-4-220	unknown	G2/M-M-G1
418 Bstc2 4-4-230		G2/M-M-G1
		G2-G2/M
	unknown	M-G1
421 Bstc3 1-1-250	unknown	G1/S-S
422 Bstc3 1-1-250		G2/M-M
	1	M-G1
46 . +	ınknown	M-G1
10-1	ınknown	M-G1
1400	ınknown	G1/S-S-G2
	inknown	G1/S-S-G2/M
428 Bstc3 -13-330	ınknown	S-G2
100 b		G1
	nknown	G2-M
1424 0 1 0 10 00	nknown	S-G2-M-G1
	nknown	G2/M-M-G1
	nknown	M-G1
46.	nknown	M-G1
40F B + 0 + 1	nknown	M-G1
	nknown	G2/M-M-G1
407 b + 0 0 + 15	nknown	G1/S; M-G1
130 0	nknown	G1/S-S-G2
	hknown	S-G2/S
440 0	known	M-G1
	known	G1/S-S-G2
	known	G1/S; G2/M-M-G1
33		_

442 Bstc3 2-2-390	L	loonana
443 Bstc3 2-2-93		G2/M-M-G1 G2/M-M
444 Bstc3 2-3-100	unknown	
445 Bstc3 -23-125	unknown unknown	S-G2 G2/M-M-G1
446 Bstc3 2-3-155	UIIKIIOWII	S-G2-M
447 Bstc3 -23-175	unknown	G2/M-M-G1
448 Bstc3 -23-177	UTIKHOVIT	G2/S-G2-M-G1
449 Bstc3 2-3-63	unknown	S-G2
450 Bstc3 -23-65	Unknown	S; G2-M-G1
451 Bstc3 -24-155	unknown	G2/M-M-G1
452 Bstc3 2-4-230	unknown	G2/M-M
453 Bstc3 2-4-250	unknown	G2/M-M-G1
454 Bstc3 -24-255	unknown	G2/M-M-G1
455 Bstc3 -24-305		G2-M-G1
456 Bstc3 -24-340	unknown	G1/S-S; M-G1
457 Bstc3 -24-90		M-G1
458 Bstc3 -31-130	unknown	G1/S-S-G2
459 Bstc3 3-120	unknown	G1/S-S
460 Bstc3 -31-200		S-G2
461 Bstc3 -31-260	unknown	G1/S-S
462 Bstc3 3-150	unknown	G2/M-M-G1
463 Bstc3 -32-105	unknown	G2-G2/M
464 Bstc3 -32-120	ļ	G1/S-S; G2/M-M-G1
465 Bstc3 -32-240	unknown	S-G2
466 Bstc3 -32-320	ļ	G1/S-S-G2; M-G1
467 Bstc3 3-280	unknown	G2-M-G1
468 Bstc3 3-2-90	unknown	S-G2
469 Bstc3 3-3-105	unknown	G2/M-M-G1
470 Bstc3 3-3-115	 	G1/S-S; M-G1
471 Bstc3 3-3-165 472 Bstc3 -34-110	 	G1/S-S-G2/S
473 Bstc3 3-4-165	 	G2/M-M G2/M-M
474 Bstc3 3-4-200	 	S
475 Bstc3 -34-290	unknown	G2/M-M-G1
476 Bstc3 -34-85	unknown	G2-M-G1
477 Bstc3 -34-90	unknown	G1/S-S
478 Bstc3 3-90	unknown	S
479 Bstc3 4-115		G2-M-G1
480 Bstc3 -41-180		G2/M-M-G1
481 Bstc3 4-13-300	unknown	G/S-S;M-G1
482 Bstc3 4-3-100		M-G1
483 Bstc3 4-3-135		S-G2-G2/M
484 Bstc3 4-3-190		S-G2-M-G1
485 Bstc3 -43-210	unknown	G1/S-S; M-G1
486 Bstc3 4-3-210		G2/S-G2-G2/M
487 Bstc3 -43-240		G1/S-S; G2/M-M-G1
488 Bstc3 4-3-248		S
		G2/M-M-G1
490 Bstc3 -43-280		G2/M-M-G1
491 Bstc3 4-3-95		S C1/S S: M C1
492 Bstc3 -44-155 493 Bstc3 -44-173		G1/S-S; M-G1
		G2/M-M-G1
495 Bstc4 -11-117		S-G2/S G2/M-M-G1
		M-G1
400		G2-M-G1
498 Bstc4 -11-180		G2/M-M-G1
400 6		G1/S-S-G2
500 ft + + +		G2/M-M-G1
		G1/S-S-G2/S
		G1/S-S-G1/S
503 Bstc4 1-1-245		M-G1
504 Bstc4 -11-350		G2/M-M
505 Bstc4 1-1-90		G2/M-M-G1
COO		G2-M-G1

		1 € 17 ₺1 05/05 / 05
507 Bstc4 1-2-280		S-G2-M
508 Bstc4 -13-112	unknown	S-G2
509 Bstc4 1-3-170	unknown	G1/S-S
510 Bstc4 1-3-205	unknown	G2/M-M-G1
511 Bstc4 -13-280	unknown	G1/S-S-G2/S
		
512 Bstc4 -13-70	unknown	G2/M-M-G1
513 Bstc4 1-4-105		M-G1
514 Bstc4 1-4-112	unknown	G2/M-M
515 Bstc4 -14-120	unknown	G1/S-S; M-G1
516 Bstc4 1-4-127	unknown	S-G2-M
517 Bstc4 1-4-145	unknown	G2/M-M-G1
518 Bstc4 -14-160	unknown	G2/M-M-G1
519 Bstc4 1-4-165	unknown	G2-M-G1
520 Bstc4 1-4-185	GITKITOWIT	G1/S-S-G2
521 Bstc4 1-4-270		
521 Bstc4 1-4-270 522 Bstc4 2-1-150	 	G1/S-S; G2/M-M-G1
	unknown	G2/M-M-G1
523 Bstc4 -21-155		G1/S-S-G2
524 Bstc4 -21-200	unknown	S; G2/M-M-G1
525 Bstc4 2-135	<u>unknown</u>	G2/M-M-G1
526 Bstc4 -22-150	unknown	G1/S-S; G1
527 Bstc 42-2-170		S-G2-M
528 Bstc4 2-2-185		M-G1
529 Bstc4 2-2-220	unknown	M-G1
530 Bstc4 2-3-100	unknown	M-G1
531 Bstc4 -23-115		
	unknown	M-G1
532 Bstc4 2-3-133		S-G2/S
533 Bstc4 -23-135	unknown	G2/M-M-G1
534 Bstc4 2-4-110	unknown	G1/S-S; G2/M-M-G1
535 Bstc4 -24-240		G1/S-S-G2
536 Bstc4 -31-260		G2/M-M-G1
537 Bstc4 -31-310	unknown	S; G2/M-M-G1
538 Bstc4 3-3-100		S-G2-M
539 Bstc4 3-3-103	unknown	G2/M-M-G1
540 Bstc4 3-3-135	- Control of the cont	M-G1
541 Bstc4 3-3-175		G2/M-M-G1
542 Bstc4 3-3-250	unknown	
	unknown	M-G1
543 Bstc4 -34-135	unknown	G2/M-M-G1
544 Bstc4 -34-185	 	G1/S-S
545 Bstc4 3-4-200	unknown	G2/M-M-G1
546 Bstc4 3-4-320		G1/S-S
547 Bstc4 -41-100	unknown	G2-M
548 Bstc4 -41-105	unknown	G1/S-S; G2/M-M-G1
549 Bstc4 -41-107	unknown	G2/M-M-G1
550 Bstc4 -41-125	unknown	M-G1
551 Bstc 4-41-180		G2/M-M-G1
552 Bstc4 -41-220	unknown	M-G1
553 Bstc4 4-150	unknown	G2-M-G1
554 Bstc4 -42-110	unknown	G2/M-M-G1
555 Bstc4 -42-115	unknown	G2/M-M
556 Bstc4 -42-130	unknown	S-G2
557 Bstc4 -42-165	unknown	G1/S-S; M-G1
558 Bstc4 -42-217	unknown	G2/M-M-G1
559 Bstc4 -43-103	unknown	G1/S-S-G2-G2/M
560 Bstc4 4-3-167	unknown	G2/M-M-G1
561 Bstc4 4-3-170		M-G1
562 Bstc4 4-4-120	unknown	M-G1
	unknown	
563 Bstc4 4-4-290	unknown	G2/M-M-G1
564 Bstt1 -11-190	 	G1/S-S
565 Bstt1 -11-200	unknown	G1/S-S-G2-G2/M
566 Bstt1 -11-55	unknown	G1/S-S
567 Bstt1 -11-65	unknown	G1/S-S-G2
568 Bstt1 -12-105	unknown	G2/M-M
569 Bstt1 -12-115	T	G1/S-S
		101/0-0
570 Bstt1 -12-230 571 Bstt1 -13-150	unknown	S-G2 G2/M-M

WO 03/085115 PCT/EP03/03703

WO 03/0851	115	
572 Bstt1 -13-230	unknown	lG2/S-G2-M
573 Bstt1 -14-125	unknown	G1/S-S
574 Bstt1 -14-220	unknown	G2/M-M
575 Bstt1 -21-100	unknown	G2/M-M
576 Bstt12 -1-240	unknown	S-G2-M
577 Bstt1 -21-250	unknown	S; G2/M-M-G1
578 Bstt12 -2-100	unknown	G2/S-G2-M-G1
579 Bstt12 -2-140	unknown	
580 Bstt1 -22-160	unknown	G2/M-M-G1
	- Indiana	G2/M-G1
581 Bstt12 -2-215	unknown	G2/M-M
582 Bstt1 -22-225	 	M-G1-G1/S
583 Bstt12 -2-360	unknown	G2/M-M-G1
584 Bstt1 -22-70	unknown	G1/S-S
585 Bstt12 -3-115	unknown	G1/S-S-G2
586 Bstt1 -23-150	unknown	G2-M-G1
587 Bstt1 -23-170	unknown	G2-M
588 Bstt12 -3-170	unknown	G1/S-S
589 Bstt1 -23-180	unknown	G2/S-G2-M
590 Bstt1 -23-185		G2-M-G1
591 Bstt1 -23-235	unknown	G2-M
592 Bstt1 -24-105	unknown	G2/S-G2-M-G1
593 Bstt1 -24-120	unknown	G2/M-M-G1
594 Bstt12 -4-260		G2/S-G2-G2/M
595 Bstt12 -4-320	1	G2/M-M
596 Bstt1 -31-120	 	G2/M-M-G1
597 Bstt1 -31-180	unknown	G2/M-M-G1
598 Bstt13 -170	unknown	
599 Bstt13 -2-150	UTIKHOWH	G1/S-S-G2
		G1/S-S-G2
600 Bstt1 -32-170	unknown	G1/S-S-G2
601 Bstt1 -32-185	 	G1/S-S
602 Bstt13 -3-100	unknown	G1/S-S-G2-M
603 Bstt1 -33-170	unknown	G1/S-S-G2
604 Bstt13 -3-320	unknown	G2/M-M-G1
605 Bstt1 -33-66		G2/M-M
606 Bstt1 -41-120	unknown	G2/M-M
607 Bstt1 -42-264	unknown	G2-M-G1
608 Bstt14 -2-280	unknown	G2/M-M-G1
609 Bstt14 -3-120		S-G2
610 Bstt14 -3-140	unknown	G1-S-S-G2
611 Bstt1 -43-220	unknown	G2/S-G2-G2/M
612 Bstt1 -43-330	unknown	G2/M-M-G1
613 Bstt14 -3-460	unknown	G2/M-M
614 Bstt14 -4-130	unknown	S-G2
615 Bstt14 -4-150	unknown	G2
616 Bstt14 -4-195	1	S-G2-M
617 Bstt14 -4-220		G2/S-G2-G2/M
618 Bstt14 -85	nohits	G2/M-M
619 Bstt21 -1-170	unknown	G2/M-M
620 Bstt2 -11-290		G2/S-G2-G2/M
621 Bstt2 -11-540	 	G1/S-S
622 Bstt21 -2-190		G2/M-M-G1
623 Bstt2 -13-165	<u> </u>	
624 Bstt2 -13-170	unknow=	S-G2-M
625 Botto 44 400	unknown	G2/M-M
625 Bstt2 -14-130	unknown	G2/M-M
626 Bstt2 -14-175	unknown	S-G2
627 Bstt22 -1-140	unknown	S-G2
628 Bstt2 -21-300	unknown	G2/M-M
629 Bstt22 -2-110	unknown	G1/S-G2
630 Bstt22 -2-255		G1/S-S-G2-G2/M
631 Bstt22 -2-370		G1/S-G2
632 Bstt22 -3-100	unknown	G2/M-M-G1
633 Bstt22 -3-145	unknown	G2/M-M-G1
634 Bstt2 -23-220	unknown	G2-M-G1
635 Bstt2 -23-370		G1/S-G2
636 Bstt22 -4-145	unknown	G2/M-M

637 Bstt22 -4-170		S-G2
638 Bstt22 -4-175		G2-M
639 Bstt22 -80	unknown	G2/M-M
640 Bstt23 -1-128	unknown	S-G2
641 Bstt23 -1-155	unknown	S-G2-G2/M
642 Bstt2 -31-200	unknown	G2/S-G2
643 Bstt23 -170	unknown	G2/M-M-G1
644 Bstt2 -32-175	unknown	G2/S-G2-G2/M
645 Bs tt23-220		G1/S-S-G2
646 Bstt23 -3-200		G1/S-S-G2/S
647 Bstt23 -3-265	-	S-G2-G2/M
648 Bstt23 -3-330		G1/S-S
649 Bstt2 -34-170		
	unknown	G2/M-M-G1
650 Bstt23 -4-180		S-G2-M
651 Bstt23 -4-210	<u> </u>	G2/M-M-G1
652 Bstt2 -41-170	unknown	G1/S-S-G2
653 Bstt24 -1-170	unknown	S-G2
654 Bstt2 -41-390		S-G2
655 Bstt2 -42-300		G2/M-M-G1
656 Bstt24 -2-318	7	S-G2
657 Bstt24 -2-320	unknown	G2/M-M-G1
658 Bstt24 -290	unknown	G2/M-M
659 Bstt2 -43-150		S-G2
660 Bstt2 -43-160	 	S-G2/S
661 Bstt2 -43-50		
		S
662 Bstt2 -43-65	unknown	S-G2
663 Bstt2 -44-230		G2/S-G2-M
664 Bstt2 -44-240	unknown	G1/S-S-G2
665 Bstt244-240	unknown	G1/S-S-G2/S
666 Bstt24 -4-260	unknown	G1/S-S
667 Bstt24 -4-283	unknown	G1/S-S-G2
668 Bstt24 -4-285	unknown	G2/M-M-G1
669 ₿stt31 -1-145		S-G2-M
670 Bstt31 -1-210		G2/M-M-G1
671 Bstt31 -2-165	unknown	G2/S-G2
672 Bstt31 -2-185		G2/M-M-G1
673 Bstt3 -12-200	unknown	G2/M-M-G1
674 Bstt3 -12-315	anna ovin	S-G2-M
675 Bstt31 -2-330	 	G2/M-M-G1
676 Bstt3 -13-110	unknown	
677 Bstt31 -3-180	unknown	S-G2-G2/M
		S-G2-G2/M
678 Bstt3 -13-360	ļ. <u>.</u>	G2/M-M
679 Bstt3 -14-130	unknown	G2/M-M
380 Bstt3 -14-135	unknown	G2/M-M
681 Bstt31 -50	unknown	G1/S-S-G2-G2/M
682 Bstt32 -1-105		S-G2
583 ₿stt3 -21-165		G2/S-G2
684 Bstt3 -21-305	unknown	G2/M-M
85 Bstt32 -140	unknown	S-G2/S
86 Bstt3 -22-100	1	G2/M-M-G1
687 Bstt32 -2-210		S-G2-M
688 Bstt3 -22-280	unknown	G1/S-S;M-G1
889 Bstt32 -2-510	unknown	S-G2-G2/M
	MIKIOWII	
390 Bstt32 -3-115	Lumber acces	G2/S-G2
91 Bstt32 -3-155	unknown	S-G2
92 Bstt32 -3-160	 	M
393 Bstt32 -3-180	unknown	G1/S-S-G2
694 Bstt3 -23-205	unknown	S-G2-M
95 Bstt3 -23-65	unknown	G2/M-M-G1
96 Bstt32 -4-170	unknown	S; M
97 Bstt32 -4-195		G1/S-S;G2/M-M-G
	unknown	G1/S-S
99 Bstt3 -24-390		M-G1
00 Bstt33 -1-105		G1/S-S-G2
'01 Bstt33 -1-128		S-G2

WO 03/085115

WO 03/0851	15	
702 Bstt33 -1-132	unknown	G2/M-M
703 Bstt33 -1-160	unknown	G2/M-M-G1
704 Bstt33 -1-185		M-G1
705 Bstt33 -140	unknown	G2/M-M-G1
706 Bstt33 -2-75	unknown	G1/S-S-G2
707 Bstt33 -2-85	di introvir	G1/S-S; G2/M-G1
	 	G1/S-S; G2/M-M-G1
708 Bstt33 -3-110		G2/M-M-G1
709 Bstt33 -3-125	unknown_	
710 Bstt3 -33-170	unknown	S-G2/S
711 Bstt33 -4-110	<u> </u>	S-G2
712 Bstt33 -4-120	unknown	G1/S-S-G2
713 Bstt33 -4-130	unknown_	G2/M-M
714 Bstt33 -95	unknown	G2/M-M
715 Bstt34 -1-110		S-G2-G2/M
716 Bstt34 -1-170		G1/S-S-G2-G2/M
717 Bstt3 -42-350	unknown	G2/M-M-G1
718 Bstt3 -43-145	unknown	G2/M-M-G1
719 Bstt3 -43-190	unknown	G1/S-S; M-G1
720 Bstt3 -43-265	UTIKTOVIT	G2/S-G2-M-G1
	unknown	G2/M-M-G1
721 Bstt3 -43-280	unknown	
722 Bstt34 -70	unknown	S -
723 Bstt41 -3-100b	unknown	G2/M-M
724 Bstt41 -3-130	unknown	G2/M-M-G1
725 Bstt41 -3-140	unknown	G2/M-M-G1
726 Bstt41 -3-180		G2-M
727 Bstt41 -3-230	unknown	S-G2
728 Bstt41 -3-90	unknown	G2/M-M-G1
729 Bstt41 -4-210	unknown	S-G2-M-G1
730 Bstt4 -14-500	1	G2/M-M-G1
731 Bstt41 -70	unknown	G1/S-S
732 Bstt42 -1-130	unknown	G2/M-M-G1
	unknown	G2/M-M
733 Bstt42 -1-290 734 Bstt4 -21-60	unknown	S-G2
	Ulikilowii	
735 Bstt4 -22-100	 	M-G1
736 Bstt4 -22-360	 	S-G2
737 Bstt42 -3-105	unknown	G1/S-S-G2/S
738 Bstt42 -3-110	unknown	G2/M-M-G1
739 Bstt4 -23-130	<u> </u>	S-G2/M
740 Bstt4 -23-160	<u> </u>	G2/S-G2-M
741 Bstt42 -4-150	unknown	G1/S-S-G2
742 Bstt4 -24-270	unknown	G2/M-M-G1
743 Bstt42 -4-390	unknown	M-G1
744 Bstt43 -1-290	unknown	G2/M-M-G1
745 Bstt43 -1-85	1	G1/S-S-G2/S
746 Bstt4 -32-230	unknown	G1/S-S-G2/S
747 Bstt43 -2-238	- CATHOTOWIT	G2/M
	unknown	G1/S-S-G2
748 Bstt43 -3-145	unknown	G2/M-M-G1
749 Bstt43 -3-210	1	
750 Bstt43 -4-230	unknown	G2/M-M-G1
751 Bstt4 -34-75	unknown	G2/S-G2-M
752 Bstt44 -1-125	unknown	S-G2-G2/M
753 Bstt44 -185	unknown	M-G1
754 Bstt44 -2-135		G2/M-M-G1
755 Bstt4 -42-150	unknown	M
756 Bstt4 -42-390	unknown	M-G1
757 Bstt44 -3-240	unknown	G2/M-M-G1
758 Bstt44 -3-250	unknown	S-G2-G2/M
759 Bstt4 -44-148		G2/M-M-G1
	unknown	G2/M-M
760 M Bc02-100	unknown	
761 M Bc02-120	unknown	G2/M-M
762 M Bc03-110	unknown	G2/M-M
763 M Bc03-85		G2/M-M
764 M Bc11-135	unknown	G2-M
765 M Bc12-150		S-G2-M
766 M D-24 405	Linknows	G2/M-M

766 M Bc31-185

unknown

PCT/EP03/03703

767 M Bc32-107	unknown	G2/M-M-G1
768 M Bc32-110	unknown	G2/M-M-G1
769 M Bc41-110	unknown	G1/S-S; G2/M-M
770 M Bc42-280	unknown	G2-M
771 M Bc43-95	unknown	G2-M
772 M Bc44-130		S-G2
773 M Bc44-95	unknown	G2/M-M
774 M Bt12-80	unknown	G2/M-M
775 M Bt12-95		M
776 M Bt13-105	unknown	M-G1
777 M Bt14-100	unknown	G2/M-M-G1
778 M Bt14-85	unknown	S-G2-M
779 M Bt14-90	unknown -	G2-M
780 M Bt31-95		S-G2-M
781 M Bt33-115		G2/M-M-G1
782 M Bt33-133		G2-M
783 M Bt42-135	unknown	G2-M
784 M Bt43-95	unknown	G2-G2/M
785 M Bt44-145	unknown	G1/S-S-G2-M

WO 03/085115 PCT/EP03/03703

CLAIMS

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1. A method for identifying and validating plant genes/proteins as targets for agrochemicals, said method comprising the steps of:

- Determining gene or protein expression profiles during a biological process of a plant or plant cell, said biological process being necessary for the growth and/or development and/or viability of the plant or plant cell;
- b. Selecting genes or proteins having altered expression during said biological process,
- c. Cloning said selected gene or the nucleic acid encoding said protein in its full-length or partial form,
- d. Incorporating said nucleic acid in a vector designed for downregulation of expression of said nucleic acid or the sequence homologous to said nucleic acid in a plant or plant cell.
- 2. The method according to claim 1, wherein said biological process cell division.
- 3. The method according to claim 1 or 2, wherein said gene or protein expression profiling is based on nucleic acid or protein samples collected from a synchronized culture of dividing plant cells.
- 4. The method according to claim 3, wherein said dividing plant cells are tobacco BY2 cells.
 - 5. The method according to any of claims 1 to 4, wherein the expression profiles are determined by means of micro-array, macro array or c-DNA-AFLP.
- 25 6. The method according to any of claims 1 to 5, wherein said downregulation involves a viral-induced gene silencing mechanism.
 - 7. The method according to any of claim 1 to 6, wherein said downregulation involves the use of infectious DNA of virus is Tobacco Rattle Virus and wherein said plant is tobacco.
 - 8. A method for screening candidate agrochemical compounds comprising the use of any of the methods according to claim 1 to 10.
- 9. A method for screening candidate agrochemical compounds comprising the use of any one or more of SEQ ID NO 1 to 785 or a homologue, functional fragment or derivative thereof or one or more of the proteins corresponding to SEQ ID NO 1 to 785 or a homologue, functional fragment or derivative thereof.

WO 03/085115 PCT/EP03/03703

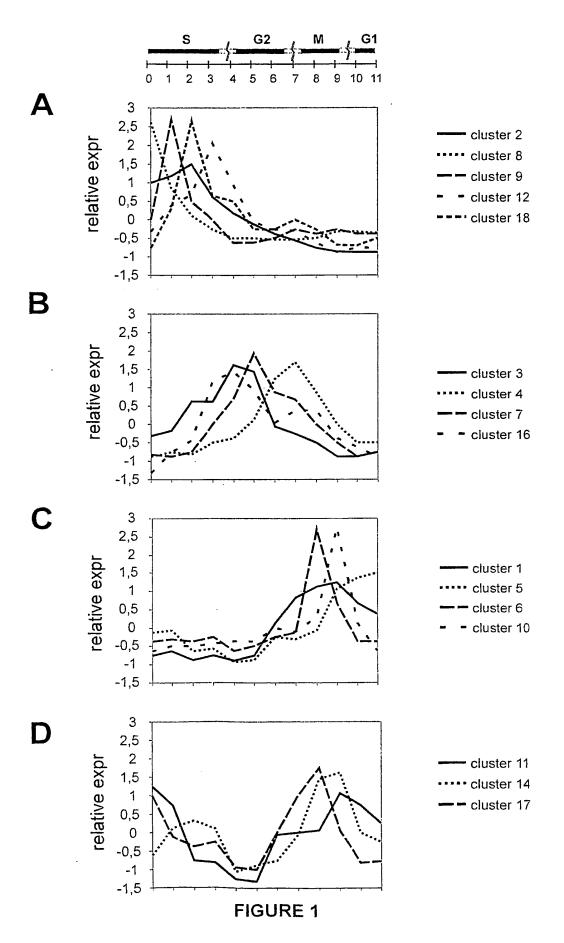
10. A method for the production of an agrochemical resistant plant, comprising the use of any one or more of SEQ ID NO 1 to 785 or a homologue, functional fragment or derivative thereof or one or more of the proteins encoded by SEQ ID NO 1 to 785 or a homologue, functional fragment or derivative thereof.

- 11. An isolated nucleic acid identifiable by any of the methods according to claims 1 to 10.
- 12. An isolated nucleic acid, comprising at least part of a nucleic acid sequence chosen from the group of SEQ ID NO 1 to 785 a homologue, functional fragment or derivative thereof.
 - 13. Use of a gene nucleic acid according to claim 11 or 12 or the protein encoded by said isolated nucleic acid as a target for an agrochemical compound.
- 15 14. Use of a nucleic acid or protein according to claim 13, wherein the agrochemical compound is a herbicide.
 - 15. A plant tolerant to an agrochemical, in which the expression level of one or more of the nucleic acids corresponding the SEQ ID NO 1 to 785 or the homologue, functional fragment or derivative thereof, is modulated.
 - 16. A harvestable part of a plant according to claim 15.

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Exp. II - 12 days after inoculation

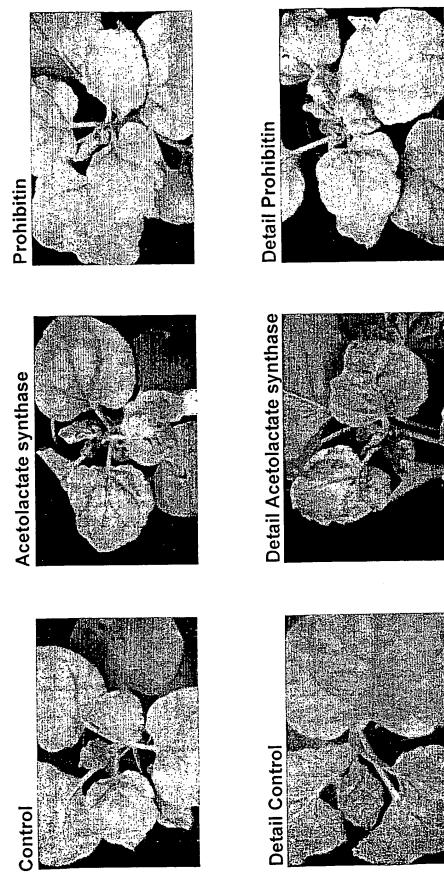


FIGURE 2

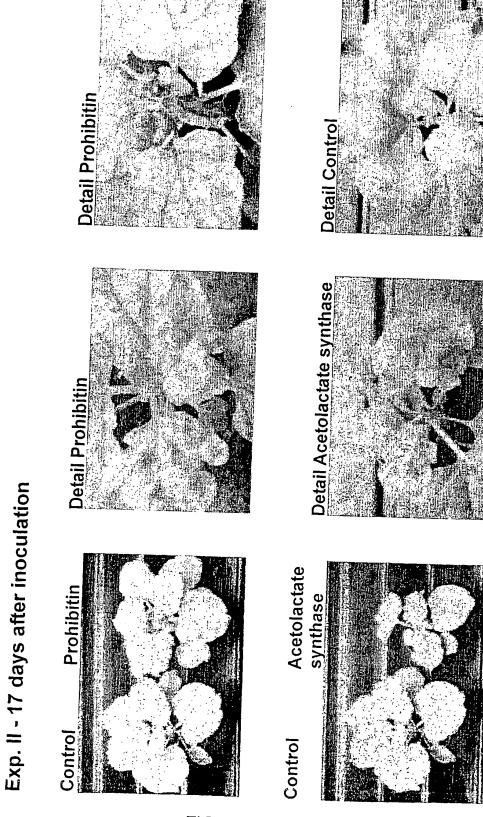
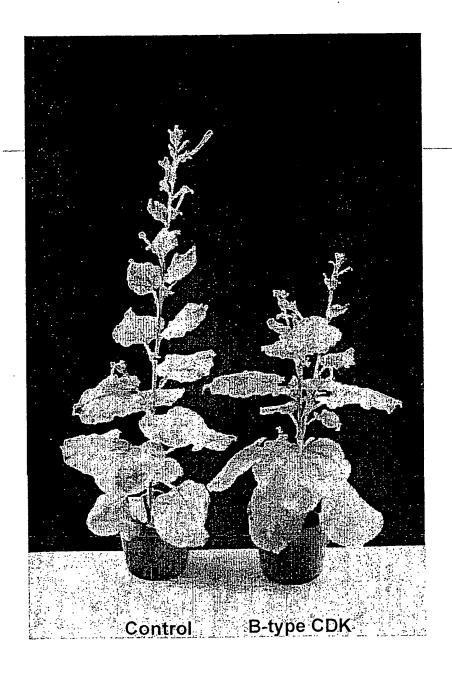


FIGURE 2 (continued)

Exp. III - 37 days after inoculation



Sequence Listing

GROUP 1

SEQIDNO1

GAATTCACTAGTGATTGATGAGTCCTGAGTAAGGTGAGACGAGAAGCGACCTTCTGACCACA AGACTTGTCAGCCTGAGACAGGTATGATATCCATATACTGCGTATCTCATAAGTGACTCGTG GATCGGATAAATGCTCAACCCATTTGCTAACATATCTGTCTTGCCTGTCAGGTTCCCAGGAT CACTACGCAGTCAATCGAATTCCCGCGGCCTATAGTGAGTCGTATTAA

SEQIDNO2

SEQIDNO3

 ${\tt GNNATGCCCGATNTAAGCCGCCCCTANATACANTTNAAATGGTCCCGGANACCCTGGGNGACAATNATNGACTGNGCAGTGGTTGAAGNTTGACAATTCCTATT}$

SEQIDNO4

SEOI DNO5

TTTTANGNCANCAAATCTCNCTCTAACGGACCCTNGCATGGCTTGTTCAAAATAAATGCCTC AGGACAATACCACGTTATGTAATGGGGAGTGAACTGCGTATATCCGTTCTGCTCNTTTATCT GGGCGGNGCCTTTGAAGTTTTTGACAAACTCTNTCTGGNTCTCACACTTAGGGCCACACTCA TCATTACTGTTTGTCCAAAACTCGTACTCAACTCTTTCATCGGGATGTGGAAGCGCCTCTCT CCAATCAAGGTTTATG

SEOIDNO6

CTTGGATGGTCNACCAGATTGAAGAACNCGAGAAAAAGCTGTTTTCTCATCCACTTCATAAG TCACAAAATGAACANCAGCCNTTGAGAATCNCAGCTGTGNTATGTANNTTCGAAGACATTGG CTGAGGATGCTGCATGGAAGTTTGTGAAAGAGAAAGCCTATCGATATGGTTACGATAAACCC AGCAATGGTTATTGGCGGTTTGTTACAACCAATAC

SEQIDNO7

GCGGTTGATATGTGGTCTGTGGGATGTATTTTTTGCCGAGATGGTTCGAAGGCAAGCCTTATT
TCCTGGTGACTCTGAGTTTCAGCAACTGCTTCACATATTCAGGCTGTTAGGAACCCCAACTG
AGAAGCAGTGGCCTGGAGTCAGTTCACTCCGCGACTGGCATGTTTATCCAAAATGGGAACCT
CAGAACTTGGCCTCTGCTGTTCCAGCATTGGGTCCTGATGGCGTGGACCTCCTCACGAAAAT
GCTCCAATATGATCCGGCAGATAGGATTTCAGCAAAAGCTGCACTTGATCATCCATACTTCG
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SEQIDNO9

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SEQIDNO10

SEQIDNO11

CGTTGNTTGTTTCGGGAAATTGGAACAGCATTGGTGAAGGCACTTACGGNCAAGTGTACATG GNTAAAGAAATTAGAACAGGGGAAATTGTTGCNTTGAAGAAGATACGCATGGACAACGAANG AGAAGGGTTTCCAATANCTGCTATACGTGAAATCAAAATCTTGAAGAAGCTGCACCATGAAA ATGNGA

SEQIDNO12

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SEOIDN013

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SEQIDNO14

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SEOIDNO15

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FIGURE ** ** *

SEOIDN016

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SEQIDNO17

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GAAGAGGGGGTTTCCAAGCTTTGCTCTGG

SEQIDNO18

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SEOIDNO19

SEQIDNO20

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SEQIDNO21

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GROUP 2

SEQIDNO22

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FIGURE 47 P

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SEOIDNO146

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SEQIDNO148

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 ${\tt GCCAGGTGCTCTCCCTAGGAAGAGTATCCTGAAATAGTTGCAGAACAGCTTTACAGGNTTCTGCAAGAGAAGTTTGAGCTTNAGGC}$

SEOIDNO150

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manner of the college

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SEQIDNO160

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SEOIDNO161

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SEQIDNO162

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SEQIDNO166

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SEQIDNO169

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SEOIDNO173

SEOIDNO174

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SEQIDNO176

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SEOIDNO177

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SEQIDNO178

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SEQIDNO179

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SEQIDNO183

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SEOIDNO184

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SEQIDNO185

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SEQIDNO186

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SEOIDNO188

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SEQIDNO189

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SEOIDNO190

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SEOIDNO191

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SEOIDNO192

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SEQIDNO193

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SEQIDNO195

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SEOIDNO196

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SEQIDNO197

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SEQIDNO198

SEOIDNO199

SEQIDNO200

SEQIDNO201

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SEQIDNO202

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SEOIDNO203

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SEQIDNO205

SEQIDNO206

SEQIDNO207

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SEQIDNO208

SEQIDNO209

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SEQIDNO210

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SEQIDNO216

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SEOIDNO221

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SEOIDNO224

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SEQIDNO229

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SEOIDNO230

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SEOIDNO231

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SEOIDNO232

SEOIDNO233

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SEQIDNO236

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SEOIDNO243

SEQIDNO244

SEQIDNO245

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PCT/EP03/03703 WO 03/085115

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Group 4

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96/140

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SEQIDNO324

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SEQIDNO349

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SEQIDNO408

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WO 03/085115 120/140

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SEQIDNO743

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SEOIDNO744

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SEQIDNO746

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SEOIDNO751

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SEQIDNO757

SEQIDNO758

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SEQIDNO768

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SEQIDNO769

SEQIDNO770

SEQIDNO771

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SEQIDNO774

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SEOIDNO775

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SEQIDNO778

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SEOIDNO788

GACGATGAGTCCTGAG

SEOIDNO789

TACTCAGGACTCAT

SEOIDNO790

GACTGCGTAGTGATCNNN

SEOIDNO791

GATGAGTCCTGAGTAANN

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(19) World Intellectual Property Organization

International Bureau



(43) International Publication Date 16 October 2003 (16.10.2003)

(10) International Publication Number WO 2003/085115 A3

(51) International Patent Classification⁷: G01N 33/53

C12N 15/82.

(21) International Application Number:

PCT/EP2003/003703

8 April 2003 (08.04.2003) (22) International Filing Date:

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

02447062.7 60/396,124

EP 10 April 2002 (10.04.2002) 15 July 2002 (15.07.2002) US

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(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NI, NO, NZ, OM, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

with international search report

(88) Date of publication of the international search report: 5 August 2004

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: IDENTIFICATION AND VALIDATION OF NOVEL TARGETS FOR AGROCHEMICALS

(57) Abstract: The invention relates to a method for identifying and validating plant targets for agrochemicals, comprising the steps of determining gene or protein expression profiles in function of the progression of an essential biological process in a plant, and the subsequent downregulation of expression of said gene or protein in a plant cell. More particularly, the effects of downregulation of the candidate target gene were directly monitored on plants locally infected with a vector mediating viral induced gene suppression in that infected plant area. The invention also relates to isolated plant genes encoding proteins involved in plant growth and development. The invention also relates to plants tolerant to agrochemicals such as herbicides or pesticides.

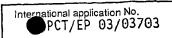


International Application No PC P 03/03703

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A. CLASSI IPC 7	FICATION OF SUBJECT MATTER C12N15/82 G01N33/53					
According to	According to International Patent Classification (IPC) or to both national classification and IPC					
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Minimum do	ocumentation searched (classification system followed by classification ${\tt C12N} - {\tt G01N}$	on symbols)				
Documental	tion searched other than minimum documentation to the extent that s	uch documents are included in	n the fields searched			
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Electronic d	ata base consulted during the international search (name of data bas	se and, where practical, searc	h terms used)			
EPO-In	ternal, BIOSIS, WPI Data, MEDLINE, I	EMBL				
C. DOCUM	ENTS CONSIDERED TO BE RELEVANT					
Category °	Citation of document, with indication, where appropriate, of the rele	evant passages	Relevant to claim No.			
Υ	US 6 369 296 B1 (BAULCOMBE DAVID ET AL) 9 April 2002 (2002-04-09)	CHARLES	1-8			
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Υ	BREYNE PETER ET AL: "Genome-wide expression analysis of plant cel	1-8				
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X Funti	ner documents are listed in the continuation of box C.	X Patent family member	ers are listed in annex.			
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"O" document referring to an oral disclosure, use, exhibition or document is combined wi			with one or more other such docu- n being obvious to a person skilled			
"" document published prior to the international filing date but		in the art. "&" document member of the	same patent family			
Date of the actual completion of the international search		Date of mailing of the international search report 2 7. 10, 2003				
1	8 August 2003	<u> </u>				
Name and n	nailing address of the ISA	Authorized officer				
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fay: (+31-70) 340-3016		Bucka, A.				

PC 17 EP 03/03703

	TO BE EVANT	PC1/EF 03/03/03
C.(Continue Category *	ation) DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 01 94603 A (ROBERTSON DOMINIQUE; TURNAGE MICHAEL A (US); UNIV NORTH CAROLINA () 13 December 2001 (2001-12-13) the whole document	1-8
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Law and in claims	s were found unsearchable (Continuation of Item 1 of first sheet)
Box I Observations where certain claims	S WOLD LOUIS ALL STATES
This International Search Report has not been esta	ablished in respect of certain claims under Article 17(2)(a) for the following reasons:
Claims Nos.: because they relate to subject matter not	required to be searched by this Authority, namely:
Claims Nos.: because they relate to parts of the Internations an extent that no meaningful Internations.	national Application that do not comply with the prescribed requirements to such al Search can be carried out, specifically:
	are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of inve	ention is lacking (Continuation of item 2 of first sheet)
)	tiple inventions in this international application, as follows:
see additional sheet	
As all required additional search fees we searchable claims.	vere timely paid by the applicant, this International Search Report covers all
As all searchable claims could be searchable claims could be searchable claims.	ched without effort justifying an additional fee, this Authority did not invite payment
As only some of the required additional covers only those claims for which feet	al search fees were timely paid by the applicant, this international Search Report s were paid, specifically claims Nos.:
4. No required additional search fees we restricted to the invention first mention See Invention 1.	ore timely paid by the applicant. Consequently, this International Search Report is need in the claims; it is covered by claims Nos.:
Remark on Protest	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

Invention 1: claims 1 to 8

a method for identifying and validating plant targets for agrochemicals

Inventions 2 to 786: claims 9 to 16 (all partially)

each invention comprises the use of one nucleic acid selected from the group of SEQ ID NO: 1-785 as a target for a herbicide or pesticide, a method of screening candidate agrochemical compounds using said nucleic acid, the use of said nucleic acid to produce agrochemical resistant plants,

the corresponding isolated nucleic acid

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WO 0107601	A	01-02-2001	US AU BR CA EP JP WO US	630384 623810 001268 238036 119655 200350507 010766 200206942	0 A 5 A 3 A1 6 A2 9 T 01 A2	16-10-2001 13-02-2001 16-04-2002 01-02-2001 17-04-2002 12-02-2003 01-02-2001 06-06-2002
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